TYK2–STAT1–BCL2 Pathway Dependence in T-cell Acute Lymphoblastic Leukemia

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ABSTRACT
Targeted molecular therapy has yielded remarkable outcomes in certain cancers, but specific therapeutic targets remain elusive for many others. As a result of two independent RNA interference (RNAi) screens, we identified pathway dependence on a member of the Janus-activated kinase (JAK) tyrosine kinase family, TYK2, and its downstream effector STAT1, in T-cell acute lymphoblastic leukemia (T-ALL). Gene knockdown experiments consistently showed TYK2 dependence in both T-ALL primary specimens and cell lines, and a small-molecule inhibitor of JAK activity induced T-ALL cell death. Activation of this TYK2–STAT1 pathway in T-ALL cell lines occurs by gain-of-function TYK2 mutations or activation of interleukin (IL)-10 receptor signaling, and this pathway mediates T-ALL cell survival through upregulation of the antiapoptotic protein BCL2. These findings indicate that in many T-ALL cases, the leukemic cells are dependent upon the TYK2–STAT1–BCL2 pathway for continued survival, supporting the development of molecular therapies targeting TYK2 and other components of this pathway.

SIGNIFICANCE: In recent years, “pathway dependence” has been revealed in specific types of human cancer, which can be important because they pinpoint proteins that are particularly vulnerable to antitumor-targeted inhibition (so-called Achilles’ heel proteins). Here, we use RNAi technology to identify a novel oncogenic pathway that involves aberrant activation of the TYK2 tyrosine kinase and its downstream substrate, STAT1, which ultimately promotes T-ALL cell survival through the upregulation of BCL2 expression. Cancer Discov; 3(5); 564–77. ©2013 AACR.

See related commentary by Fontan and Melnick, p. 494.
The prosurvival members of the BCL2 family as well as pathways that signal upstream of these proteins are attractive candidate targets in T-ALL, as these proteins are known to determine whether developing T cells undergo apoptosis in the thymus or survive to reach peripheral organs (12, 13). Normally, thymocytes will only survive to maturity if they can productively rearrange their T-cell receptors (TCR) such that they react with foreign antigens and do not react with self-antigens. In contrast, the vast majority of thymocytes that fail to rearrange their TCRs in this manner are eliminated by activation of proapoptotic BCL2 family members followed by caspase-mediated cell death. Defective signaling through this pathway would enable thymocytes slated for destruction to survive and acquire additional lesions that promote full malignant transformation. This suggests that T-ALL cells may have acquired a dependence on this specific pathway whose actions perturb the normal balance between thymocyte life and death signaling cues.

Here, we identify pathway dependence in T-ALL on the aberrant activation of tyrosine kinase 2 (TYK2), a member of the prosurvival members of the BCL2 family as well as pathways that signal upstream of these proteins are attractive candidate targets in T-ALL, as these proteins are known to determine whether developing T cells undergo apoptosis in the thymus or survive to reach peripheral organs (12, 13). Normally, thymocytes will only survive to maturity if they can productively rearrange their T-cell receptors (TCR) such that they react with foreign antigens and do not react with self-antigens. In contrast, the vast majority of thymocytes that fail to rearrange their TCRs in this manner are eliminated by activation of proapoptotic BCL2 family members followed by caspase-mediated cell death. Defective signaling through this pathway would enable thymocytes slated for destruction to survive and acquire additional lesions that promote full malignant transformation. This suggests that T-ALL cells may have acquired a dependence on this specific pathway whose actions perturb the normal balance between thymocyte life and death signaling cues.

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the Janus-activated kinase (JAK) family that phosphorylates and activates STAT1 and leads to the upregulation of BCL2, which is then required for T-ALL cell survival.

**RESULTS**

**Loss-of-Function RNAi Screens**

To understand the oncogenic contribution of tyrosine kinases in T-ALL, we conducted an RNA interference (RNAi)-assisted protein target identification (RAPID) screen of primary leukemic cells from a pediatric patient with T-ALL, applying validated siRNAs to silence each member of the tyrosine kinase family (14). The result showed clear dependence of these leukemic cells on the TYK2 tyrosine kinase for their viability (Fig. 1A and Supplementary Table S1). We independently conducted an RNAi screen in which 5,000 inducible short-hairpin RNAs (shRNA) targeting 1,740 genes (15, 16) were introduced into 3 T-ALL cell lines (JURKAT, CCRF-CEM, and SKW-3/KE-37). By determining the relative abundance of each shRNA in shRNA-induced versus uninduced samples after 3 weeks of induction, we identified shRNAs that were significantly depleted in T-ALL cell lines (Supplementary Table S2). Notably, an shRNA targeting TYK2 was depleted from cultures of these T-ALL cell lines (Fig. 1B), indicating that this gene is required for T-ALL cell survival or proliferation, whereas control diffuse large B-cell lymphoma cells showed little to no depletion of cells harboring TYK2-specific shRNA.

We then asked whether this activity was restricted to TYK2 or if it might be shared with other members of the JAK family tyrosine kinases. By testing multiple lentiviral shRNA constructs targeting JAK1, JAK2, JAK3, or TYK2 in JURKAT cells (Supplementary Fig. S1 and Supplementary Table S3), we found that knockdown of TYK2, but not of other JAK family genes, reduced cell growth (Fig. 1C), indicating that the cells specifically depend on TYK2 within the JAK family kinases. Significantly, we observed growth inhibition for each of the 3 independent shRNAs in a long-term culture (Fig. 1D). After analyzing additional T-ALL cell lines, we found that the growth of 14 (88%) of 16 successfully transduced cell lines was significantly inhibited after silencing of TYK2 (“TYK2-dependent” cells), whereas 2 others (LOUCY and TALL-1) were unaffected (“TYK2-independent” cells; Supplementary Table S4). Representative results with 5 cell lines are shown in Fig. 1E. To further explore TYK2 dependence in primary T-ALL specimens, we transfected TYK2 or control siRNA ex vivo into leukemic cells derived from patients with T-ALL that had been directly expanded in immunocompromised mice (“primagraft” samples). We found that 5 (62.5%) of 8 primagrma samples (#1–5) showed sensitivity to silencing of TYK2 (Fig. 1F). These results indicate that a significant fraction of T-ALL cells from human patients depend on TYK2 for sustained growth.

To determine the mechanism(s) underlying the impaired cell growth observed after TYK2 silencing, we stained cells with Annexin V and found increased percentages of apoptotic cells in the TYK2-dependent cell lines JURKAT, RPMI-8402, and HPB-ALL, but not in the TYK2-independent cell line LOUCY (Fig. 1G). TYK2 knockdown had little effect on cell-cycle distribution (Supplementary Fig. S2), indicating that the T-ALL cells primarily require TYK2 for survival rather than proliferation. Expression of the cDNA containing only the coding region of wild-type TYK2 rescued the JURKAT cells from apoptosis (Fig. 1H, left, as indicated by PARP cleavage) and growth inhibition (Fig. 1H, right), confirming that shRNA targeting TYK2 induces apoptosis due to its silencing of TYK2 and not to any off-target effects.

**TYK2 Pathway Includes STAT1 and BCL2**

On the basis of our results implicating TYK2 signaling as essential for T-ALL cell survival, we investigated downstream mediators of the TYK2 pathway. STAT1 is an attractive candidate effector of TYK2 signaling, as STAT1 is phosphorylated on tyrosine and activated downstream of TYK2 activation in cytokine signaling pathways such as the interleukin (IL)-10 pathway (Fig. 2A; ref. 17). Indeed, tyrosine phosphorylation of STAT1 was observed in 16 of 19 T-ALL cell lines, most of which were TYK2 dependent (Supplementary Table S4), whereas tyrosine phosphorylation of STAT2, for example, was not observed (Supplementary Fig. S3A). STAT1 was also phosphorylated in the LOUCY and TALL-1 TYK2-independent cell lines, presumably due to activation by other pathways, indicating that these cell lines do not depend on the TYK2 pathway for survival. Further investigation showed that silencing of endogenous TYK2 resulted in decreased STAT1 phosphorylation in JURKAT cells, whereas overexpression of wild-type TYK2 caused increased STAT1 phosphorylation (Fig. 2B). We also observed constitutive phosphorylation of STAT3 in many T-ALL cell lines (Supplementary Fig. S3), although knockdown or overexpression of TYK2 did not change the phosphorylation status of STAT3 (Fig. 2B). STAT5 phosphorylation has been associated with other genetic abnormalities, such as mutation of IL-7R (DND-41; ref. 18), NUP214–ABL fusion (ALL–S1L and PEER; ref. 19), and LCK–TCR fusion (H-SB2; ref. 20), whereas AKT phosphorylation has been mostly associated with deletion/mutation of the PTEN gene (21). Phosphorylation of extracellular signal–regulated kinase (ERK) 1/2 was detected only in some cell lines. Importantly, knockdown of STAT1 decreased the growth of each of 2 TYK2-dependent cell lines (JURKAT and HPB-ALL) but not that of the LOUCY TYK2-independent cell line (Fig. 2C and Supplementary Fig. S4A). Thus, STAT1 seems to be a component of the TYK2-mediated pathway that promotes cell survival in T-ALL. This activity stands in marked contrast to type 1 IFN signaling through TYK2, which leads to tyrosine phosphorylation of both STAT1 and STAT2 (Supplementary Fig. S3B).

To identify the effector molecules downstream of the TYK2–STAT1 pathway in T-ALL, we analyzed global gene expression profiles in JURKAT cells after silencing of TYK2 or STAT1. Gene set enrichment analysis (GSEA) revealed that a shared set of genes was downregulated after either TYK2 or STAT1 silencing (Fig. 2D). Importantly, we found that mRNA encoding the BCL2 antiapoptotic protein was significantly downregulated after silencing of both TYK2 and STAT1 (Fig. 2E). Other antiapoptotic BCL2 family proteins and inhibitor of apoptosis (IAP) family proteins were not significantly downregulated after silencing of TYK2 or STAT1, and proapoptotic BCL2 family proteins were not upregulated after TYK2 knockdown (Supplementary Table S5). Analysis
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A, primary cells from a pediatric patient with T-ALL were electroporated with siRNAs targeting each member of the tyrosine kinome as well as N-RAS, K-RAS, and control scrambled siRNA. Cell viability values are shown in relation to the group mean ± 2 SDs. Values above or below the 2-SD threshold were considered significant.

B, shRNA screen with approximately 5,000 inducible shRNAs that collectively target 1,740 genes were conducted on 3 T-ALL cell lines (JURKA T, CCRF-CEM, and SKW-3/KE-37) and 4 diffuse large B-cell lymphoma (DLBCL) cell lines (L3, Ly10, Ly7, and Ly19). Depletion of TYK2 shRNA from the cell population was calculated as shRNA-uninduced/induced (log2), and is shown as the mean ± SEM of 4 independent experiments.

C, validated shRNAs targeting JAK1, JAK2, JAK3, or TYK2 as well as 2 control shRNAs (GFP and Luc) were transduced by lentivirus infection into JURKA T cells. The number of shRNAs tested is indicated in parentheses. Cell viability was measured after 3 and 7 days of infection. Growth rate (day 7/day 3) relative to control is shown as the mean ± SEM (n = 3-6). * P < 0.05 by 2-sample, 2-tailed t test.

D, the 3 TYK2-targeting shRNAs as well as control GFP shRNA were transduced by lentivirus infection into JURKA T cells. Relative cell growth values (means ± SEM of triplicate experiments) at days 3, 5, 7, and 9 after infection are shown. The 3 TYK2-targeting shRNAs as well as control GFP shRNA were transduced in 5 T-ALL cell lines (JURKA T, RPMI-8402, HPB-ALL, MOLT-4, and LOUCY). Growth rate (day 7/day 3) relative to control is shown as the mean ± SEM of triplicate experiments.

E, primary T-ALL cells were initially expanded by primagraft into RAG2−/−c−/− or NOG/Scid/IL2γ−/− mice, subsequently expanded on OP9-DL1 or MS5-DL1 cells, and electroporated with nonspecific siRNA (control) or siRNA targeting TYK2 followed by a 4-day culture. Values represent mean percent cell viability (normalized to viability of control siRNA) ± SEM of quadruplicate experiments.

F, primary T-ALL cells were then transduced by lentivirus infection with control GFP or TYK2#2 shRNA, which targets the 3′ untranslated region of TYK2 mRNA. Whole-cell extracts were harvested and subjected to immunoblot analysis with antibodies specific for total TYK2, PARP, and α-tubulin (internal control). Growth rate (day 7/day 3) relative to control is shown as the mean ± SEM of triplicate experiments. *** P < 0.001 by 2-sample, 2-tailed t test.
TYK2–STAT1 pathway upregulates the antiapoptotic protein BCL2 in T-ALL. A, diagram of proposed TYK2 pathway. B, JURKAT cells expressing an empty vector or wild-type (WT) TYK2 cDNA were transduced with TYK2 or control GFP shRNA. Whole-cell extracts were harvested and subjected to immunoblot analysis with antibodies specific for total TYK2, phospho-STAT1 (Y701), STAT1, phospho-STAT3 (Y705), STAT3, and α-tubulin. C, the 2 STAT1–targeting shRNAs as well as control GFP shRNA were transduced into JURKAT, HPB-ALL, and LOUCY cells. Cell viability was measured after 3 and 7 days of infection. Growth rate (day 7/day 3) relative to control is shown as the mean ± SEM of triplicate experiments. *, P < 0.05; ***, P < 0.001 by 2-sample, 2-tailed t test.

D, global gene expression in JURKAT cells transduced with TYK2, STAT1, or control shRNAs (GFP and Luc) was measured by microarray analysis. The genes significantly downregulated by TYK2 or STAT1 knockdown (KD) compared with control were determined and used as gene sets for the GSEA. GSEA plots indicate the degree to which genes are overrepresented at the extreme left (downregulated by KD) or right (upregulated by KD) of the entire ranked list. Solid bars represent genes. Normalized enrichment score (NES) and P values are indicated. E, BCL2 mRNA expression levels in each shRNA-transduced sample were determined by microarray analysis. Values are means ± SEM of duplicate experiments. ***, P < 0.001 by 2-sample, 2-tailed t test. F, the TYK2 shRNA as well as control GFP shRNA were transduced into T-ALL cell lines. BCL2 mRNA expression was measured by quantitative RT-PCR and normalized by glyceraldehyde-3-phosphate dehydrogenase (GAPDH) expression. Gene expression changes (KD/control) were shown as mean ± SEM of duplicate experiments. G, the WT or kinase-dead form of TYK2 cDNA was transduced into MOLT-4 cells by retrovirus infection. The selected clone was then transduced with GFP or TYK2 shRNA by lentivirus infection. Whole-cell extracts were subjected to immunoblot analysis with antibodies specific for TYK2, BCL2, and α-tubulin. H, apoptosis was measured after 4 days of infection by flow cytometric analysis of cells stained with Annexin V–Phycocerythrin (PE). Values are means ± SEM of duplicate experiments. ***, P < 0.001 by 2-sample, 2-tailed t test. I, the WT or Y701F STAT1α cDNA was transduced in JURKAT cells by retrovirus infection. The selected clone was transduced with GFP or STAT1 shRNA by lentivirus infection. Whole-cell extracts were subjected to immunoblot analysis with antibodies specific for total STAT1, phospho-STAT1 (Y701), BCL2, and α-tubulin. Arrowhead indicates STAT1α isoform. J, apoptosis was measured after 4 days of infection by flow cytometric analysis of cells stained with Annexin V–PE. Values are means ± SEM of duplicate experiments. *, P < 0.05 by 2-sample, 2-tailed t test.
by quantitative PCR of additional T-ALL cell lines revealed that silencing of TYK2 resulted in significant reductions of BCL2 mRNA expression in multiple TYK2-dependent cell lines, but not in the LOUCY TYK2-independent T-ALL cell line (Fig. 2F), which seems to have constitutively upregulated BCL2 through a pathway independent of TYK2. Expression of wild-type but not kinase-dead TYK2 (M978F) protein was sufficient to rescue BCL2 protein expression and to prevent apoptosis after knockdown of endogenous TYK2 in the TYK2-dependent line (Fig. 2G and H). In fact, expression of the kinase-dead TYK2 protein repressed BCL2 expression (Fig. 2G) and increased baseline levels of apoptosis (Fig. 2H), suggesting that it could be interfering with residual endogenous TYK2 activity. Similarly, expression of the shRNA-resistant wild-type STAT1α protein partially rescued BCL2 protein expression (Fig. 2I) and prevented apoptosis (Fig. 2J), whereas STAT1α Y701F (which is incapable of being activated by phosphorylation) did not rescue BCL2 levels and caused increased levels of apoptosis. Taken together, these results show that TYK2 effects are in part mediated through STAT1, and that the TYK2–STAT1 pathway acts at least in part by upregulating BCL2 expression in T-ALL cells, thereby promoting their aberrant survival.

Activating TYK2 Mutations in T-ALL

To investigate the possibility that the TYK2 kinase might be mutationally activated, we sequenced the TYK2 coding regions in 17 T-ALL cell lines and 45 primary pediatric T-ALL patient samples (Supplementary Tables S6 and S7). This analysis identified novel nonsynonymous sequence variants of TYK2 in 4 cell lines (21.1%), none of which represents known single-nucleotide polymorphisms based on the dbSNP137 and the 1000 Genomes databases (22, 23). These included point mutations in the TYK2 FERM domain (G36D, S47N and R425H), the JH2 domain (V731I), and the kinase domain (E957D and R1027H; Fig. 3A and 3B and Supplementary Table S6). The functional consequence of each point mutation was then determined by testing its ability to transform IL-3-dependent murine pro-B cells into B-lineage cells, thereby promoting their survival.

Variants harboring point mutations in the TYK2 FERM domain (G36D, S47N, and R425H) and the JH2 domain (V731I) and the kinase domain (E957D) and R1027H) were endogenously expressed in mouse T-lineage cells in murine bone marrow stromal cells plated on OP9 DL1 stromal cells. Transduction of TYK2-E957D together with GFP into bone marrow cells significantly fostered cell growth compared with the result for cells expressing an empty vector or wild-type TYK2 (Fig. 3G); flow cytometric analysis of the expanded cells confirmed the expression of both CD3 and GFP (Supplementary Fig. S5).

TYK2 Pathway Activation by IL-10 Signaling

Mutations of the TYK2 gene provide a mechanism for aberrant TYK2 activation in some T-ALL cell lines; however, many T-ALL samples that were TYK2 dependent lacked mutations within this gene, implicating an alternative mechanism(s) of kinase activation that drive pathway dependence. In the context of cytokine signaling, TYK2 is known to be activated by upstream receptors, including the type 1 IFN (IFNα/β), IL-6, IL-10, IL-11, IL-12, IL-23, and IL-27 receptors (17, 26). Thus, we hypothesized that autocrine activation of one or more of these receptors might be responsible for activating the TYK2 pathway. Among the known cytokine pathways, autocrine IL-10 receptor signaling was an attractive possibility because IL-10 has been reported to increase BCL2 expression and cell survival in hematopoietic progenitor cells (27), primary T cells (28), and germinal center B-cells (29). To test this notion, we evaluated the effects on cell growth of shRNAs specific for the IL-10 and for IL-10 receptor genes (IL10RA and IL10RB; Supplementary Fig. S4A), which are endogenously expressed in all cell lines examined (Supplementary Fig. S6A and S6B). In 2 T-ALL cell lines that are TYK2-dependent but lacking a mutation of the TYK2 gene (HPB-ALL and JURKAT), knockdown of IL10, IL10RA, or IL10RB resulted in a reduction of cell growth (Fig. 4A) and the induction of apoptosis (Fig. 4B), coincident with downregulation of BCL2 expression (Fig. 4C), indicating that IL-10 signaling is required for T-ALL cell survival. Of note, knockdown of other cytokine receptor genes (IFNAR1, IFNAR2, IL6ST, IL11RA, IL12RB1, IL12RB2, and IL27RA) did not downregulate BCL2 expression in T-ALL cells (Supplementary Fig. S4B). IL6R and IL23R, which encode other cytokine receptors that can interact with TYK2, were not expressed in T-ALL cell lines.

Knockdown of the IL-10 receptor genes in these cell lines resulted in downregulation of STAT1 phosphorylation (Fig. 4D), indicating that the IL-10 receptor is required for activation of the TYK2–STAT1 pathway. Treatment of these cell lines with an anti-IL-10–neutralizing antibody did not show any effects on cell growth (Supplementary Fig. S6C), suggesting that this pathway may be activated during intracellular trafficking of the ligand and receptor. In contrast, little or no effect on cell growth or survival was observed when these shRNAs were introduced into the LOUCY TYK2-independent cell line. In MOLT-4 cells that harbor an activating TYK2 mutation (E957D), knockdown of the IL-10 receptor genes resulted in lower levels of BCL2 and induced apoptosis, suggesting that the IL-10 receptor proteins may still be required as a scaffold when the TYK2 protein is mutationally activated.
To assess the efficacy of JAK inhibitors for impairing the growth and viability of TYK2-dependent T-ALL cells, we chose 3 compounds (JAK inhibitor I, AG490, and CP-690550) with differing degrees of efficacy against TYK2 compared with other JAK family members. JAK Inhibitor I has been shown to exhibit potent activity against all JAK family kinases, including TYK2 (30). Indeed, Ba/F3 cells transformed by constitutively active JAK kinases (TYK2-E957D, TEL-JAK1, TEL-JAK2, and TEL-JAK3) were consistently sensitive to this inhibitor compared with a negative control Ba/F3 cell line transformed by an alternative tyrosine kinase, TEL-ABL (Fig. 5A). Further use of JAK inhibitor I against T-ALL cell lines revealed that the TYK2-dependent cell lines were sensitive to this inhibitor.
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**Figure 4.** IL-10 receptor signaling is required for TYK2 activation and cell survival in T-ALL. **A,** validated shRNAs targeting IL10RA, IL10RB, or IL10 as well as control GFP shRNA were transduced by lentivirus infection into 4 T-ALL cell lines (HPB-ALL, JURKAT, MOLT-4, and LOUCY). Relative cell growth at days 3, 5, 7, and 9 after infection was evaluated. Values are means ± SEM of triplicate experiments. **B,** apoptosis was measured after 6 days of infection by flow cytometric analysis of cells stained with Annexin V–FITC. Values are means ± SEM of duplicate experiments. *P < 0.01; ***P < 0.001 by 2-sample, 2-tailed t test. **C,** BCL2 mRNA expression level was measured by reverse transcription PCR (RT-PCR) and normalized by glyceraldehyde-3-phosphate dehydrogenase (GAPDH) expression. BCL2 expression relative to control is shown as the mean ± SEM of duplicate experiments. **D,** whole-cell extracts harvested from HPB-ALL or JURKAT cells transduced with shRNAs targeting control GFP, IL10RA, or IL10RB were subjected to immunoblot analysis with antibodies specific for total STAT1, phospho-STAT1 (Y701), and α-tubulin.

with IC_{50} values ranging from 1–3 μmol/L, whereas a TYK2-independent cell line, LOUCY, was insensitive (Fig. 5B). In the JURKAT TYK2-dependent cells, the cell number was significantly lower in the inhibitor-treated sample than in the control after long-term treatment, whereas this effect was less pronounced in the LOUCY TYK2-independent line (Fig. 5C). A broader analysis of 19 T-ALL cell lines revealed a strong correlation between sensitivity to TYK2 shRNA and sensitivity to JAK kinase inhibitors (Supplementary Table S4). Treatment with AG490, a pan-JAK family inhibitor (31), yielded essentially the same results as JAK inhibitor I (Supplementary Fig. S7A). In contrast, T-ALL cell lines treated with CP-690550, a potent inhibitor of JAK2 and JAK3 but not TYK2 (32), were resistant to this compound (Supplementary Fig. S7B). These results indicate that the sensitivity of T-ALL cells to JAK inhibitor I and AG490 is not due to inhibition of JAK2 or JAK3, but rather to activity against TYK2, consistent with our initial findings using RNAi gene silencing techniques (Fig. 1A–C). Immunoblots of cell lysates after treatment with JAK inhibitor I showed decreased phosphorylation of both TYK2 and STAT1 in the Ba/F3 cells expressing TYK2-E957D (Fig. 5D). Moreover, apoptosis was induced in TYK2-dependent lines after JAK inhibitor I treatment, as indicated by increased levels of Annexin V staining, which was not observed in the TYK2-independent LOUCY cells (Fig. 5E). Flow cytometric analysis of TYK2-dependent lines after treatment with JAK inhibitor I showed an increase in the number of sub-G_{1} cells, consistent with ongoing apoptosis (Supplementary Fig. S8).

We also observed an increase of G_{2}–M fraction cells; however, these changes in the cell cycle were not apparent by shRNA TYK2 knockdown in the same cell lines (Supplementary Fig. S2). Further investigation will be required to determine whether this is a specific effect of inhibiting TYK2. Taken together, these results indicate that pharmacologic inhibition of TYK2 kinase activity induces the death of TYK2-dependent T-ALL cells.

**DISCUSSION**

Our studies establish dependence on the TYK2–STAT1 pathway in 14 of 16 T-ALL cell lines and 5 of 8 primagraft samples. These findings are the first to implicate TYK2 activation in oncogenesis and to show that the oncogenic TYK2 signaling pathway differs from previously described TYK2 signaling pathways, in which TYK2 and JAK1, as well as other JAK kinases, crossactivate one another. Porcu and colleagues (33) reported that the HPB-ALL TYK2-dependent cell line expresses a different JAK1 transcript variant, which does not encode a critical portion of the kinase domain (33), suggesting selective pressure to inactivate the JAK1 kinase in T-ALL. Two other TYK2-dependent cell lines, MOLT-4 and RPMI-8402, express another JAK1 transcript variant that generates a truncated form of JAK1 protein lacking the entire kinase domain. Thus, inactivation of JAK1 kinase through mutation or alternative splicing may contribute to the noncanonical TYK2-mediated pathway that promotes cell survival.
Figure 5. Activity of small-molecule inhibitors of TYK2 against transformed Ba/F3 and T-ALL cells. A, Ba/F3 cells transformed by TEL–ABL, TEL–JAK1, TEL–JAK2, TEL–JAK3, or TYK2–E957D and cultured with graded concentrations of JAK Inhibitor I for 3 days. Cell viability values are means ± SEM percentage of the untreated control values in triplicate experiments. B, LOUCY, JURKAT, MOLT-4, HPB-ALL, and RPMI-8402 cells cultured with graded concentrations of JAK Inhibitor I for 3 days. Cell viability values are means ± SEM percentages of the untreated control in 6 experiments. C, JURKAT and LOUCY cells were cultured in the absence or presence of JAK inhibitor I (3 μmol/L) for 7 days. The cell number is shown as fold change from day 0 for each cell line (means ± SEM of duplicate experiments). D, Ba/F3 cells expressing TYK2-E957D cultured with graded concentrations of JAK inhibitor I and subjected to immunoblot analysis with antibodies specific for phospho-TYK2, phospho-STAT1, PARP, and α-tubulin. E, JURKAT, HPB-ALL, and LOUCY cells cultured with graded concentrations of JAK inhibitor I for 24 hours, and assessed for apoptosis by flow cytometric analysis after staining with Annexin V-FITC. Values are means ± SEM percentages for duplicate experiments. *, P < 0.05; ***, P < 0.001 by 2-sample, 2-tailed t test.
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in T-ALL. Alternatively, activating mutations of JAK1 have also been observed in some cases of T-ALL, mostly in adult patients—1 of 49 (2%; ref. 34), 4 of 108 (4%; ref. 35), 8 of 38 (21%; ref. 34), and 3 of 11 (27%; ref. 36). Hence, although activated JAK1 may contribute to T-ALL pathogenesis in some cases, in other T-ALL cases, reliance on TYK2 signaling for cell survival, there seems to be concomitant selective pressure to inactivate signaling molecules downstream of JAK1.

We found that the TYK2–STAT1 pathway positively regulates BCL2 expression in T-ALL cells, contributing to aberrant cell survival. It has been reported that mitochondria from chemosensitive cancer cells such as ALL are consistently more "primed," so that the cells are more prone to undergo apoptosis than are those from normal tissues (37). Lymphoma cells require BCL2 in addition to other antiapoptotic BCL2 family members to sequester proapoptotic proteins such as BIM, indicating that inhibition of BCL2 protein is a useful strategy to induce apoptosis in already "primed" leukemia cells. The fact that the ABDT-737 BCL2 antagonist induces apoptosis in BCL2-overexpressing chronic lymphoid leuke-

mia cells (38) supports this finding. Our results are consistent with the dependence of T-ALL cells that are "primed" to undergo apoptosis on upregulated levels of BCL2 expression induced by TYK2, and this model explains why the T-ALL cells die when TYK2 is silenced and BCL2 levels fall. BCL2 is normally not detectable in immature thymocytes in the thymic cortex but is markedly upregulated in more mature single-positive T-cells that migrate into the thymic medulla (12, 39), thereby enabling functionally selected thymocytes to survive. This antiapoptotic protein is also frequently overexpressed in T-ALL as well as in T-cell lymphoblastic lymphoma (T-LBL), a distinct clinical presentation of thymocyte (41), and there is no difference in BCL2 protein expression in the thymus between Tyk2 knockout and wild-type mice (Supplementary Fig. S9). Thus, TYK2 is not required for the upregulation of BCL2 expression by normal medullary thymocytes, but its aberrant gain-of-function activity in immature double-positive T-ALL/T-LBL cells can nevertheless promote BCL2 overexpression and aberrant cell survival, leading to clonal expansion of fully transformed leukemic lymphoblasts.

Our work indicates that in a subset of T-ALLs lacking TYK2-activating mutations, the TYK2 prosurvival pathway could be activated downstream of autocrine activation of IL-10 and its receptors. IL-10 is a multifunctional cytokine with pleiotropic effects on different hematopoietic cell types (42). IL-10 signaling is primarily involved in limiting or terminating inflammatory responses, at least in part, by promoting the activity of regulatory T-cells. In other contexts, IL-10 has been shown to enhance the survival of hematopoietic stem cells, B-cells, T-cells, and tumor cells (27–29) through its ability to upregulate BCL2. The IL-10 ligand–receptor interaction normally engages both JAK1 and TYK2, which are constitu-

tively associated with IL-10RA and IL-10RB, respectively. In T-ALLs that depend on TYK2 activity for cell survival, we determined that JAK1 activity is not required (Fig. 1A–C), and in some of these cases, JAK1 is actually inactivated by mutation or alternative splicing (33), suggesting a noncanonical independent role for TYK2 in the aberrant survival of T-ALL cells.

We hypothesize that autocrine expression of IL-10 or, rarely, TYK2-activating mutations is acquired during thymocyte transformation, presumably at the point when mitochondria have become "primed" due to cooperating oncogenic mutations so that the cells are prone to undergo apoptosis (37). Our study has identified dependence on TYK2 kinase activity as a requirement for the survival of a substantial fraction of human T-ALL cell lines and primafagra T-ALL samples. This discovery adds a novel mechanism to the repertoire by which cancer cells block pathways leading to cell death in particular lineages. Indeed, the promotion of tumor cell survival is now an established step in the molecular pathogenesis of human cancers, including leukemias, which is distinct from changes leading to the initiation, repopulation, self-renewal, or proliferation of transformed cells. Unless apoptosis is blocked, and the emerging precurser clone is able to survive, it will not be possible for the evolving malignant cells to select for additional mutations and acquire a fully transformed phenotype.

Our study also shows that pharmacologic inhibition of TYK2 kinase activity induces the death of TYK2-dependent T-ALL cells, which, combined with TYK2 knockdown experiments, indicates dependence on the TYK2–STAT1 pathway. A clear implication of these results is that drugs able to potently inhibit TYK2 are likely to provide a therapeutic advantage in patients with T-ALL. A loss-of-function mutation in the TYK2 gene was identified in a patient with hyperimmunoglobulin E syndrome (43), a primary immunodeficiency characterized by elevated serum immunoglobulin E, which in this case was associated with an increased susceptibility to infection by various microorganisms. These findings support the notion that TYK2 inhibition would not have a significant effect on the development of normal T cells or these precursors or hematopoietic stem/progenitor cells, although it might cause susceptibility to viral infections after prolonged exposure. The only effective inhibitor of TYK2 that we have identified is JAK inhibitor I, a "tool" compound that remains suboptimal for use in patients. Unfortunately, the JAK kinase inhibitors now in clinical use were developed against alternative JAK kinases, so they are not potent inhibitors of TYK2. Hence, there is a need to identify potent new drugs with tolerable side effects that could be used to target TYK2 in T-ALL cells dependent on this kinase.

METHODS

Collection of Primary T-ALL Specimens

For RAPID assay, T-ALL diagnostic specimens were collected with informed consent and Institutional Review Board (IRB) approval of Oregon Health & Science University (Portland, OR). Bone marrow cells from patients were separated on a Ficoll gradient, and mono-
nuclear cells were treated with ammonium-chloride-potassium lysis buffer. For TYK2 genotyping, T-ALL diagnostic specimens were collected with informed consent and IRB approval of Dana-Farber Cancer Institute (Boston, MA; clinical trial 00-001) and Children’s Oncology Group (Boston, MA; clinical trial 9404; Clinicaltrials.gov identi-

fier: NCT00165178). For the primagraf experiment, diagnostic T-ALL patient samples were obtained with informed consent and IRB approval of Dana-Farber Cancer Institute study 05-01, BC Cancer Agency (BCCA; British Columbia, Canada) study H06-00028, and were used with...
IRB approvals by the University of California, San Diego (UCSD; San Diego, CA) Human Research Protections Program and BCCA.

**Primagrafts**

Mouse studies were carried out in strict accordance with the recommendations of the Institutional Animal Care and Use Committee, and the protocols were approved by the Committee at the UCSD (Protocol S06015) and the University of British Columbia (Protocol A09-0771). T-ALL diagnostic specimens were transplanted into Rag2−/− mice or nonobese diabetic (NOD)/Scid/Il2rg−/− (NSG) mice to propagate the cells as “primagrafts.” Mononuclear cells were purified by Ficoll-Hypaque centrifugation before fluorescence-activated cell sorting (FACS) analysis and viable cryopreservation in liquid nitrogen. Approximately 50,000 human CD34+ cells selected from above T-ALL samples with the aid of FACSaria II (BD Biosciences) or immunomagnetic beads (Milteny Biotech) were transplanted intraperitoneally into Rag2−/− mice within 48 hours of birth. Mice were sacrificed at 8 weeks after transplantation. Bone marrow, spleen, thymus, liver, and peripheral blood were collected for FACS analysis of human CD45, CD34, CD2, and CD7 expression. Serial transplantation was done by intravenally injecting 50,000 human CD34+ cells selected from the T-ALL primary engrafted bone marrow or thymus into the neonatal Rag2−/− mice to propagate the cells. NSG primagrafts performed at BCCA were carried out by injection of 1–2 × 10^6 unsorted patient bone marrow cells either intravenously or immunomagnetic beads (Miltenyi Biotec) were transplanted intraperitoneally into Rag2−/−/Il2rg−/− neonatal mice within 48 hours of birth. Mice were sacrificed at 8 weeks after transplantation. Bone marrow, spleen, thymus, liver, and peripheral blood were collected for FACS analysis of human CD45, CD34, CD2, and CD7 expression. Serial transplantation was done by intravenally injecting 50,000 human CD34+ cells selected from the T-ALL primary engrafted bone marrow or thymus into the neonatal Rag2−/− mice to propagate the cells. NSG primagrafts performed at BCCA were carried out by injection of 1–2 × 10^6 unsorted patient bone marrow cells either intraperitoneally (tail vein) or intraperitoneally into sublethally irradiated adult recipient NSG mice. Human leukemia cells were harvested from bone marrow and spleen of morbid animals and characterized by FACS and/or TCry HDA clonality assay, as previously described (44).

**Cell Culture**

All T-ALL cell lines were stocked in our laboratory and were confirmed by DNA fingerprinting using the PowerPlex 1.2 system (Promega) in January 2013. HEK293T and Ba/F3 cells were recently obtained from American Type Culture Collection. HEK293T17 cells were kindly provided by Dr. Richard Van Etten (Tufts-New England Medical Center, Boston, Massachusetts). Ba/F3 derivatives expressing various oncogenic fusion kinases, namely, TEL-JAK1, TEL-JAK2, TEL-JAK3 and TEL-ABL, were obtained from Dr. Richard Moriggl and were described previously (45). T-ALL cells were maintained in “R10” (RPMI-1640 medium supplemented with 10% FBS, L-glutamine, penicillin/streptomycin; Invitrogen. Ba/F3 cells were maintained in 100 μL of siPORT buffer (Ambion). The RAPID screen was conducted as previously described (14, 46). Cells were aliquoted at 42 μL per well of culture media [RPMI-1640 (Ambion) and 2 μL of siRNA at 20 μg/mL per well onto a 96-well electroporation [Ambion] and 2 μL of siRNA at 20 μg/mL were added to each well (tyrosine kinase library purchased from Dharmacon/Thermo Fisher Scientific). Single and pooled nonspecific siRNA as well as siRNA against EPHAS, EPHAS, SRMS, AATK, LMTK1, N-RAS, and K-RAS (all from Dharmacon) were added separately because they are not included in the tyrosine kinase library. Cells were electroporated at 1,110 V (equivalent of 150 V per well), 200 μsec, 2 pulses, and 50,000 cells per well were replated into triplicate plates containing 100 μL per well of culture media [RPMI-1640 supplemented with 20% FBS, L-glutamine, penicillin/streptomycin, fungizone, ×1 insulin-transferrin-sodium selenite (Invitrogen), and 10−4 mol/L 2-mercaptoethanol (Sigma-Aldrich)]. For determination of cell viability, cells were subjected to the CellTiter 96 AQ One solution cell proliferation assay (MTS; Promega). All values were normalized to the median value on the plate.

**Inducible shRNA Screen**

The inducible shRNA screen was conducted as previously described (15, 16). Briefly, T-ALL cell lines were first transfected with an ecotropic retrovirus receptor. The host cells were then transduced with the tetracycline repressor gene and separated into single clones. Ecotropic retrovirus library was made by cotransfection of 1,000 retrovirus constructs into HEK293T cells with pHIT/ E6× 3* and pHIT6 plasmids. Each T-ALL cell line was infected with each of 5 retrovirus pools (1,000 shRNAs/pool) separately in biologic quadruplicate, selected by puromycin, divided into 2 groups, and cultured with or without doxycycline for 3 weeks to induce shRNA. Genomic DNAs were then harvested and bar-code sequences amplified by PCR. Samples from the shRNA-induced cells and the matched uninduced cells were labeled with Cy3 or Cy5, respectively, and hybridized onto the microarray chip for quantification of their relative abundance in each population. P values were determined by paired t test and adjusted for multiple comparisons; shRNA depletion (uninduced/induced) was then calculated for each shRNA. The shRNAs that are significantly depleted [adjusted P < 0.1, and shRNA depletion 20.585 (log2)] in T-ALL cell lines are selected (Supplementary Table S2).

**Individual shRNA and siRNA Knockdown Analysis**

All shRNA constructs cloned into the lentiviral vector pLKO.1-puro were obtained from the NCMR Consortium (Broad Institute, Cambridge, MA). Each construct was cotransfected into HEK293T cells with packaging plasmid delta 8.9 and envelope plasmid VSV-G using FuGENE 6 reagent (Roche). Supernatants containing the lentivirus were collected and filtered through a 0.45-μm cellulose acetate membrane filter. T-ALL cells were infected with lentivirus in the presence of polybrene (8 μg/mL) and HEPEs (10 mmol/L) by centrifugation at 2,500 rpm for 1.5 hours at 30°C, and the infected cells were selected by puromycin for at least 36 hours. For siRNA primagraft studies, primagraft cells that were expanded in Rag2−/− or NSG mice and subsequently on OP9-DL1 or MDS-DL1 coculture were freshly harvested, washed with PBS, and incubated with nonspecific (control) or TYK2 siRNA in siPORT buffer (Ambion). Cells were electroporated at 200 V, 0.2 μsec for 2 pulses, and then replated in culture media [RPMI-1640 supplemented with 20% FBS, L-glutamine, penicillin/streptomycin, fungizone, ×1 insulin-transferrin-sodium selenite (Invitrogen), and 10−4 mol/L 2-mercaptoethanol (Sigma-Aldrich)].

**Cell Viability, Apoptosis Assays, and Reagents**

For siRNA knockdown experiment, cells were subjected to the CellTiter 96 AQ One solution cell proliferation assay (Promega). All values were normalized to the mean of control siRNA cell viability values for each sample. For the shRNA knockdown experiment, cells were subjected to the CellTiter Glo assay (Promega). For drug treatment, cells were incubated in the presence of graded doses of JAK inhibitor I (EMD Biosciences) for 72 hours. The number of viable cells was determined with the CellTiter 96 AQ One solution cell proliferation assay. For determination of factor-independent growth, Ba/F3 cells were washed 3 times in R10, and 1 million cells were seeded into triplicate flasks in 2 mL of R10. Total viable cells were determined every day for 16 days using propidium iodide exclusion on a Guava cell counter (Guava Technologies). To analyze apoptosis, we washed the cells with PBS, incubated them in staining buffer containing fluoroscein isothiocyanate (FITC)-conjugated anti-Annexin V antibody (MBL International), and analyzed them by BD FACS Calibur (BD Biosciences). For the rescue study, we used a PE-conjugated Annexin V antibody (MBL International) because
the cells had been transduced with GFP as a selection marker for generation of a cell line that overexpresses TYK2.

**Immunoblotting**

T-ALL cell lines were lysed in radioimmunoprecipitation assay buffer [50 mmol/L Tris (pH 8.0), 10 mmol/L EDTA, 150 mmol/L NaCl, 0.5% deoxycholic acid, 0.1% SDS, 1% NP-40] with phosphatase inhibitors (1 mmol/L sodium pyrophosphate, 1 mmol/L sodium orthovanadate) and protease inhibitors [protease inhibitor cocktail (Roche) and 1 mmol/L phenylmethylsulfonyl fluoride]. Ba/F3 and HEK293T17 cells were lysed in 1× lysis buffer (Cell Signaling Tech). Equivalent amounts of protein were diluted in sample buffer (75 mmol/L Tris pH 6.8, 3% SDS, 15% glycerol, 8% 2-mercaptoethanol, 0.1% bromophenol blue) and separated by SDS-PAGE. Proteins were transferred to polyvinylidene difluoride membranes (Millipore) and subjected to immunoblot analysis with antibodies specific for TYK2, phospho-TYK2 (Y1054/1055), STAT1, phospho-STAT1 (Y701), STAT3, phospho-STAT3 (Y705), STAT5, phospho-STAT5 (Y694), ERK1/2, phospho-ERK1/2 (T202/Y204), PARP, α-tubulin (Cell Signaling Tech), or β-actin (Millipore).

**Microarray Gene Expression Analysis and Gene Set Enrichment Analysis**

JURKAT cells were transduced with control shRNA or shRNA targeting TYK2 or STAT1 in biologic duplicate. Total RNA was harvested by TRIzol followed by a column purification using the RNasy Mini kit (Qiagen). A total of 12 RNA samples (4 controls, 4 TYK2 knockdown, 4 STAT1 knockdown) were used for microarray expression analysis. Genome-wide RNA expression analysis was conducted by HG U133 plus 2.0 microarray chip (Affymetrix) at the Dana-Farber Cancer Institute. Expression data can be found at [http://www.ncbi.nlm.nih.gov/geo/](http://www.ncbi.nlm.nih.gov/geo/) under accession number GSE44652. After normalization using dChip (Dana-Farber Cancer Institute; ref. 47), 30,988 probes with presence-call more than 33% (4/12 arrays) were filtered. The genes differentially expressed between 4 control and 4 knockdown samples were selected on the basis of a lower 90% confidence interval (CI). The discovery rates by 1,000 permutations were 1.1% (for mean difference more than 50, by which the 90th percentile false discovery rate was calculated). The genes differentially expressed were subjected to immunoblot analysis with antibodies specific for TYK2 and STAT1.

**Cloning of Mutant TYK2 Genes and Creation of Stable Ba/F3 Clones**

For cloning of TYK2, cDNA was purchased from Origene and cloned into the EcoRI site of MSCV-ires-GFP (MIG). Point mutations were introduced with the Quikchange XL-II mutagenesis kit (Stratagene/Agilent Tech). Retrovirus expressing TYK2-WT, TYK2-V15A, TYK2-G36D, TYK2-S47N, TYK2-R245H, TYK2-V317I, TYK2-E957D, TYK2-R1027H, TYK2-E957D/M978Y, or TYK2-E957D/M978F was propagated in HEK293T17 cells by cotransfection of each respective cDNA construct with the EcoPak plasmid (kindly provided by Dr. Richard Van Etten) using FUGENE 6 (Roche). One millilitre of viral supernatant was mixed with polybrene (5 μg/mL), HEPES (7.5 mmol/L), and 1× 106 Ba/F3 cells and plated in a centrifuge at 2,500 rpm for 1.5 hours at 30°C. GFP-positive cells were sorted on a FACSaria (BD Biosciences) after 48 hours of infection.

**Transient TYK2 Expression in HEK293T17 Cells**

Constructs (MSCV-ires-GFP) expressing TYK2-E957D, TYK2-E957D/M978Y, or TYK2-E957D/M978F were transfected into HEK293T17 cells using FUGENE 6 (Roche). After 48 hours, cells were lysed and whole-cell extracts were subjected to immunoblot analysis as described above.

**Primary T-lineage Outgrowth Assay**

Murine bone marrow cells were harvested from 6-week-old C57BL/6 mice (The Jackson Laboratory) and infected with 2 rounds of Mig retrovirus expressing empty vector, TYK2-WT, or TYK2-E957D as previously described (48). GFP-positive cells were sorted on a FACSaria (BD Biosciences) and plated into triplicate wells of 24-well plates (105 cells per well) that had been seeded with OP9-DL1 cells (kindly provided by Dr. Juan-Carlos Zúñiga-Pflücker, University of Toronto, Sunnybrook Research Institute, Toronto, Ontario, Canada). Cells were cocultured in α-minimum essential media supplemented with 20% FBS, L-glutamine, penicillin/streptomycin, IL-7 (10 ng/mL), and FLT3 ligand (10 ng/mL; Peprotech). Viable cells from each well were counted every 2 days by PI exclusion on a Guava cell counter. Wells containing only OP9-DL1 cells without bone marrow cells were also counted, and these numbers were subtracted from total viable cells in coculture wells (most OP9-DL1 cells could be gated out by forward scatter). All numbers were normalized to the cell counts obtained after 2 days of culture to control for minor differences in seeding density of cells. At the end of the experiment, outgrowth cells were stained with an antibody specific for CD3 (BD Biosciences) and analyzed on a FACSaria (BD Biosciences).

**Sequencing Analyses**

Genomic DNA from patient samples and cell lines was prepared by DNasey (Qiagen) and used to sequence the TY2 gene (Genewiz) using previously described primers (49).
Statistical Analyses
For RAPID functional screens, we calculated the mean cell viability for all data points. Values that were greater than 2 SDs from the mean were considered significant. For cell proliferation and viability assays, a difference in treatment, dose, or time point compared with the relevant control treatment or the no-drug control was determined by analysis with a Student t test. IC50 values with a small-molecule inhibitor for each cell line were calculated by nonlinear regression using GraphPad Prism software.

Disclosure of Potential Conflicts of Interest
A.G. Fleischman has received honoraria for serving on the speakers’ bureau for Incyte. No potential conflicts of interest were disclosed by the other authors.

Authors’ Contributions
Writing, review, and/or revision of the manuscript: T. Sandra, J.W. Tyner, J. Glover, B.H. Chang, A.G. Fleischman, M. Kleppe, D. Neuber, R. Moriggl, M. Muller, N.S. Gray, C.H.M. Jamieson, L.M. Staudt, B.J. Druker
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