Efficacy of Intermittent Combined RAF and MEK Inhibition in a Patient with Concurrent BRAF- and NRAS-Mutant Malignancies


ABSTRACT

Vemurafenib, a RAF inhibitor, extends survival in patients with BRAFV600E-mutant melanoma but activates extracellular signal-regulated kinase (ERK) signaling in RAS-mutant cells. In a patient with a BRAFV600E-mutant melanoma responding to vemurafenib, we observed accelerated progression of a previously unrecognized NRAS-mutant leukemia. We hypothesized that combining vemurafenib with a MAP–ERK kinase (MEK) inhibitor would inhibit ERK activation in the melanoma and prevent ERK activation by vemurafenib in the leukemia, and thus suppress both malignancies. We demonstrate that intermittent administration of vemurafenib led to a near-complete remission of the melanoma, and the addition of the MEK inhibitor cobimetinib (GDC-0973) caused suppression of vemurafenib-induced leukemic proliferation and ERK activation. Antimelanoma and antileukemia responses have been maintained for nearly 20 months, as documented by serial measurements of tumor-derived DNA in plasma in addition to conventional radiographic and clinical assessments of response. These data support testing of intermittent ERK pathway inhibition in the therapy for both RAS-mutant leukemia and BRAF-mutant melanoma.

SIGNIFICANCE: We show that in a patient with simultaneous RAS-mutant leukemia and BRAF-mutant melanoma, intermittent RAF inhibitor therapy induced a near-complete melanoma response, and addition of a MEK inhibitor prevented RAF inhibitor-induced activation of the RAS-mutant leukemia. Intermittent therapy may permit greater pathway inhibition with less toxicity, avoid chronic relief of pathway feedback, and have enhanced effectiveness compared with chronic administration.

INTRODUCTION

Activating mutations at the V600 codon of BRAF are found in 40% to 60% of melanomas. These mutations lead to hyperactivation of the extracellular signal-regulated kinase (ERK) pathway, which causes feedback inhibition of RAS activation and maintains the RAF kinases in a monomeric state. Currently available ATP-competitive RAF inhibitors, such as vemurafenib, bind to BRAFV600E monomer and thus
Combined RAF–MEK Inhibition in RAS- and BRAF-Mutant Malignancies

inhibit its catalytic activity and activation of ERK signaling. Vemurafenib leads to clinically significant responses in nearly half of patients with BRAFV600E/K-mutated melanoma and improves progression-free and overall survival (1). This led to U.S. Food and Drug Administration (FDA) approval of vemurafenib in 2011. In contrast, in cells with sufficient levels of RAS activation, RAF forms activated dimers. Binding of vemurafenib and other RAF inhibitors to one member of the dimer pair results in transactivation of the other RAF molecule and causes activation of ERK signaling (2–4). This may stimulate proliferation of tumors with active RAS.

We previously reported a patient with metastatic BRAFV600K-mutant melanoma who, when treated with vemurafenib, experienced dramatic shrinkage of his melanoma but induction of proliferation of a previously unsuspected chronic myelomonocytic leukemia (CMML) that harbored an oncogenic NRASG12R mutation (5). In vitro, vemurafenib induced proliferation of the CMML cells, which could be blocked by concurrent MAP–ERK kinase (MEK) inhibition. We hypothesized that treating this patient with combined therapy with RAF and MEK inhibitors would treat the melanoma and reduce proliferation of the patient’s concurrent CMML.

Here, we report that combined therapy with vemurafenib and the MEK inhibitor GDC-0973 (now called cobimetinib) did indeed prevent proliferation of the CMML while maintaining a near-complete response of BRAFV600K-mutated melanoma. This was achieved and maintained with intermittent dosing of both drugs.

RESULTS

Clinical Case

The patient is a 76-year-old man with stage IV (T3aNxM1b) BRAFV600K-mutant melanoma who was started on therapy with vemurafenib in February 2012 (5). After 2 weeks of treatment, there was already a marked improvement in his melanoma, but his white blood cell (WBC) count increased to 80.9 × 10^9/L. Bone marrow evaluation revealed that the patient also had CMML harboring an NRASG12R mutation. As the CMML regressed quickly upon vemurafenib discontinuation, the patient was subsequently treated with vemurafenib at a dose of 720 mg twice daily on an intermittent dosing schedule and experienced a near-complete melanoma response. Specifically, vemurafenib was held when the patient developed toxicities (fatigue or joint pain) or when the WBC count approached 80 × 10^9/L. Once the toxicities resolved and the WBC count decreased to <20 × 10^9/L, vemurafenib was resumed. The patient was managed with intermittent dosing of single-agent vemurafenib for 49 weeks (Fig. 1A), achieving a near-complete melanoma response.

In November 2012, Genentech agreed to supply cobimetinib under a single-patient use Investigator New Drug application. Cobimetinib is a selective, noncompetitive inhibitor of MEK1/2 that inhibits ERK1/2 phosphorylation in cells and proliferation at IC_{50} values of 1.8 and 8 nmol/L, respectively (6). The patient was started on the recommended phase II dose for cobimetinib of 60 mg orally daily for 3 weeks, followed by 1 week off the drug. Vemurafenib (720 mg orally twice a day) was administered concurrently with cobimetinib. As shown in Fig. 1A, the addition of the MEK inhibitor resulted in suppression of the CMML, and the patient achieved normal WBC counts for the first time since beginning vemurafenib. The patient initially tolerated 3 weeks of vemurafenib/cobimetinib combination therapy, but subsequently experienced fatigue and anemia that required dose adjustments of both drugs. We found that 40 mg/d of cobimetinib was sufficient to inhibit vemurafenib-induced proliferation of the CMML but that 20 mg/d was not sufficient (Fig. 1A). We ultimately found that 7- to 10-day courses of combination vemurafenib (480 mg twice a day) and cobimetinib (40 mg/d) were tolerable and were not associated with elevation of the peripheral WBC count. Drug holidays of 2 to 3 weeks (or longer) were given as needed for the resolution of adverse symptoms, largely fatigue.

Control of the peripheral WBC and monocyte counts correlated with a decrease in spleen length as assessed by computed tomography (CT) scan (Supplementary Fig. S1). Bone marrow examinations performed at weeks 57 and 85 while on combination therapy showed persistent CMML without evidence of disease progression. The patient has now been on intermittent treatment for 85 weeks; for the first 50 weeks he received only vemurafenib and for the next 35 weeks he received vemurafenib with cobimetinib. The effect on the melanoma, as assessed by CT scans, is shown in Fig. 1B and Supplementary Fig. S2A–S2C. By week 40, the patient experienced a near-complete response with only a residual 1.1-cm subcarinal lymph node observed on imaging, which has been maintained despite subsequent intermittent treatment with only RAF and MEK inhibitors.

Quantitative Assessment of Circulating Tumor DNA

Antimelanoma and anti-CMML effects were also assessed by quantifying circulating tumor-derived DNA using a digital PCR assay (7, 8). The effect of treatment on melanoma-derived (BRAFV600K) and CMML-derived (NRASG12R) DNA in the plasma throughout the treatment is shown in Fig. 1C. This methodology allows for the detection and quantification of mutant alleles with sensitivity as low as 0.001% (see Methods and Supplementary Fig. S3A and S3B). The effect of combined RAF and MEK inhibition on the levels of melanoma-derived BRAFV600K throughout the treatment was consistent with tumor volume, as assessed by radiographic evaluation shown in Fig. 1B (Supplementary Fig. S2A–S2C). In fact, once the maximal radiographic antimelanoma response was achieved, circulating melanoma-derived BRAFV600K DNA was no longer detectable. Likewise, the level of CMML-derived NRASG12R DNA in the plasma correlated with the peripheral WBC count (Fig. 1C); however, CMML-derived DNA remained detectable throughout the treatment despite normal peripheral WBC counts, consistent with the observation that CMML remained detectable in the bone marrow.

Antileukemic Efficacy of MEK Inhibition in RAS-Mutant Leukemias

Given the effect of combined RAF–MEK inhibition on the patient’s monocytosis and NRASG12R-mutant allele burden, we next assessed the effects of vemurafenib and combined vemurafenib plus cobimetinib on ERK signaling in leukemic cells throughout the treatment. We measured the levels of phosphorylated ERK (pERK) in the CD14+ cells from the peripheral blood of the patient using phosphoprotein
flow cytometry. As expected and previously shown (5), activated ERK was increased in monocytes during the time of vemurafenib monotherapy as compared with the same cell population before vemurafenib therapy (Supplementary Fig. S4A). There was also an increase in the frequency of CD14+ cells in the peripheral blood at this time. In contrast, monocytes analyzed from the time of combined vemurafenib plus cobimetinib had reduced ERK activation compared with the same cells during a period of observation following vemurafenib withdrawal (Supplementary Fig. S4B). Moreover, the frequency of CD14+ cells among peripheral blood mononuclear cells was lowest in the samples obtained from the time of combined vemurafenib plus cobimetinib therapy.

We next sought to compare the efficacy of pharmacologic inhibition of MEK versus inhibition of RAF and combined RAF–MEK inhibition in myeloid leukemia cell lines that were BRAF WT/NRAS WT or BRAF WT/NRAS G12D mutant. Treatment with PLX4720 alone increased ERK phosphorylation in all of these BRAF wild-type cell lines and had minimal efficacy in inducing cell death, as expected (Fig. 2A–C). In contrast,
Combined RAF-MEK Inhibition in RAS- and BRAF-Mutant Malignancies

**RESEARCH BRIEF**

**Figure 2.** Viability curves and immunoblots revealing viability and activation of mitogen-activated protein kinase (MAPK) signaling intermediates in NRAS WT and NRAS G12D AML cell lines (indicated with *) treated with various concentrations of cobimetinib, PLX4720, or both. **A,** IC50 values of BRAF WT /NRAS G12D and BRAF WT /NRAS WT AML cell lines with exposure to cobimetinib or PLX4720. **B,** the cell viability of BRAF WT /NRAS G12D AML cell lines with exposure to varying concentrations of cobimetinib, PLX4720, or both. DMSO, dimethyl sulfoxide. In **A** and **B,** data were derived from 72 hours of exposure to cells followed by CellTiter-Glo Luminescence assessment. **C,** immunoblots of pERK1/2 and total ERK1/2 (top) in NRAS G12D-mutant and NRAS WT human leukemia cell lines exposed to vehicle, cobimetinib, PLX4720, or PLX4720 + cobimetinib. Quantification of pERK1/2 relative to total ERK1/2 is displayed at the bottom. Cells were treated with 500 nmol/L of each drug for 24 hours.
although MEK inhibition with cobimetinib had a modest and variable inhibitory effect on the NRAS wild-type cell lines (IC_{50} 2.3–13 μmol/L), the NRAS-mutant lines were highly sensitive to MEK inhibition (IC_{50} 0.66–0.94 μmol/L; Fig. 2A). Moreover, combined MEK and RAF inhibition was able to suppress NRAS-mutant leukemic proliferation. ERK phosphorylation was greater in three of four cell lines treated with combined RAF and MEK inhibition compared with MEK inhibition alone (Fig. 2C), suggesting that combined use of vemurafenib attenuated the antileukemic effect of cobimetinib.

**Genetic Analysis of the Leukemia and Melanoma**

Given the unique history of this patient harboring two simultaneous malignancies, we sought to define genetically both cancers in more detail, identify any potential genetic modifiers of response to RAF and MEK inhibition, and identify the development of new genetic events in the leukemia. The patient’s melanoma and the CMML were subjected to targeted sequencing of 279 genes known to be recurrently mutated across many malignancies using a bait-capture, next-generation sequencing assay, and custom-designed probes generated using the Nimblegen SeqCap system, essentially as previously described (Supplementary Table S1; ref. 9). The melanoma was sequenced from biopsy material before treatment with vemurafenib, whereas the CMML was sequenced from bone marrow mononuclear cells (BM MNC) before treatment with vemurafenib and also before introduction of cobimetinib.

In the melanoma, 28 somatic mutations in 20 separate genes were identified (Table 1 and Supplementary Table S2; DNA from buccal epithelia cells was used as paired normal). Twenty-three of 28 mutations (82%) involved a C → T or G → A transitions, indicative of UV damage. Of note, there were no mutations or copy-number alterations in p16INK4a or genetic changes suggestive of PI3K–AKT pathway activation that could activate a parallel signaling pathway [other than a PREX2A (10) somatic missense mutation of unknown importance]. In the CMML, genetic analysis of DNA from BM MNCs just after the initiation of vemurafenib and after introduction of cobimetinib consistently detected six mutations in four genes (NRAS, EZH2, TET2, and IDH2), all of which have been previously described in this disorder (11). Five of these mutations (83%) were C → T or G → A transitions. As previously observed by prior investigators, neither of the driver mutations (BRAF^{V600E} or NRAS^{G12R}) had a UV damage signature, raising the possibility that the driver mutations arose by non-UV mechanisms.

**DISCUSSION**

In this patient, the simultaneous presence of an NRAS-mutant leukemia in the setting of metastatic BRAF-mutant melanoma provided a dramatic example of the effects of activation of ERK by vemurafenib in RAS-mutant cells. Tumor induction has been noted in patients treated with RAF inhibitors, but most cases have been squamous cell cancers of the skin which are easily managed. In our case, acceleration of a previously undiagnosed mutant NRAS CMML occurred. Both the melanoma and the CMML were well controlled with intermittent inhibition of ERK signaling. In the first year, intermittent treatment with the RAF inhibitor alone reduced the melanoma to a near-complete response while mitigating the drug’s proliferative effects on the leukemia. Here, we describe that intermittent combined therapy with MEK and RAF inhibitors counteracted the paradoxical ERK activation induced by the RAF inhibitor in the CMML cells, leading to effective therapy for both malignancies.

The improved efficacy of combining inhibition of RAF and MEK in BRAF^{V600E}-mutant metastatic melanoma has been previously described in a phase I/II trial of dabrafenib and trametinib. Within this trial, 162 patients were randomized to

<table>
<thead>
<tr>
<th>Gene symbol</th>
<th>Amino acid change</th>
<th>Tumor variant frequency</th>
<th>Buccal epithelium variant frequency</th>
</tr>
</thead>
<tbody>
<tr>
<td>Melanoma</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>EPHB1</td>
<td>p.Q627K</td>
<td>0.49153</td>
<td>0.065</td>
</tr>
<tr>
<td>NOTCH4</td>
<td>p.W306*</td>
<td>0.10588</td>
<td>0.00291</td>
</tr>
<tr>
<td>MYB</td>
<td>p.P273S</td>
<td>0.16</td>
<td>0</td>
</tr>
<tr>
<td>BRAF</td>
<td>p.V600K</td>
<td>0.41551</td>
<td>0.00291</td>
</tr>
<tr>
<td>PREX2</td>
<td>p.D1072N</td>
<td>0.13889</td>
<td>0.00271</td>
</tr>
<tr>
<td>PTPRD</td>
<td>p.G1819R</td>
<td>0.17842</td>
<td>0</td>
</tr>
<tr>
<td>PTPRD</td>
<td>p.G1001R</td>
<td>0.13208</td>
<td>0</td>
</tr>
<tr>
<td>NOTCH3</td>
<td>p.G285E</td>
<td>0.16185</td>
<td>0</td>
</tr>
<tr>
<td>MLL2</td>
<td>p.T698L</td>
<td>0.18919</td>
<td>0</td>
</tr>
<tr>
<td>FLT1</td>
<td>p.G1086E</td>
<td>0.15175</td>
<td>0</td>
</tr>
<tr>
<td>DICER1</td>
<td>p.J445fs</td>
<td>0.2598</td>
<td>0</td>
</tr>
<tr>
<td>CDH11</td>
<td>p.A342T</td>
<td>0.21212</td>
<td>0</td>
</tr>
<tr>
<td>KEAP1</td>
<td>p.P549L</td>
<td>0.19764</td>
<td>0.0027</td>
</tr>
<tr>
<td>NOTCH3</td>
<td>p.S2262F</td>
<td>0.14976</td>
<td>0</td>
</tr>
<tr>
<td>NOTCH3</td>
<td>p.G824D</td>
<td>0.19421</td>
<td>0.00297</td>
</tr>
<tr>
<td>NOTCH3</td>
<td>p.P609S</td>
<td>0.20567</td>
<td>0</td>
</tr>
<tr>
<td>MEF2B</td>
<td>p.G290S</td>
<td>0.12745</td>
<td>0</td>
</tr>
<tr>
<td>CBLC</td>
<td>p.G312E</td>
<td>0.16667</td>
<td>0</td>
</tr>
<tr>
<td>PAK7</td>
<td>p.E518K</td>
<td>0.18779</td>
<td>0</td>
</tr>
<tr>
<td>PAK7</td>
<td>p.P424S</td>
<td>0.15909</td>
<td>0</td>
</tr>
<tr>
<td>ERG</td>
<td>p.M109I</td>
<td>0.16814</td>
<td>0</td>
</tr>
<tr>
<td>Leukemia</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>NRAS</td>
<td>p.G12R</td>
<td>0.49153</td>
<td>0.065</td>
</tr>
<tr>
<td>EZH2</td>
<td>p.R63*</td>
<td>0.45905</td>
<td>0.09259</td>
</tr>
<tr>
<td>IDH2</td>
<td>p.R140Q</td>
<td>0.06089</td>
<td>0</td>
</tr>
<tr>
<td>TET2</td>
<td>p.R1465*</td>
<td>0.439</td>
<td>0.084</td>
</tr>
<tr>
<td>TET2</td>
<td>p.Q1553*</td>
<td>0.49</td>
<td>0.123</td>
</tr>
</tbody>
</table>

* Mutational analysis performed using IMPACT assay, a high-throughput next-generation sequencing assay of 279 genes known to be mutated in cancer. Melanoma tumor biopsy, bone marrow aspirate mononuclear cells from 2 time points in the patient’s course, and DNA from buccal epithelia cells were sequenced.
Combined RAF–MEK Inhibition in RAS- and BRAF-Mutant Malignancies

A TaqMan assay was designed to amplify the region of interest (BRAFV600E or NRASG12R) and distinguish between the wild-type and mutant target using a pair of competitive fluorophore-labeled probes (available upon request). PCR reaction mixtures (20 μL), containing a limited template dilution, were partitioned into approximately 20,000 1-nL droplets using the Bio-Rad QX200 droplet generator system according to the manufacturer’s specifications. The limited template dilution ensured that the estimated number of templates per partition (λ) was within the dynamic range of the instrument. The Poisson correction factor was applied at the analysis stage to account for the eventuality of multiple template occupancy. Following the endpoint PCR amplification, carried under conducive conditions, the individual droplets were analyzed using the Bio-Rad QX200 Droplet Reader system and the proprietary analysis software QuantaLife. The mutant to wild-type ratio and the 95% confidence interval (CI) were calculated as previously described (17). To determine the limit of detection for the designed assays, we measured the lowest detectable amount of mutant target within a large excess of wild-type genomic DNA (gDNA). The observed number of mutant target copies was in close linear relationship with the expected quantities (R² > 0.99), and a single copy of mutant DNA resolved within 10² copies of wild-type DNA (Supplementary Fig. S3).

Leukemia Cell Line Drug Studies

Cell lines were originally obtained from DSMZ or the American Type Culture Collection, and all cell lines were authenticated by Promega short-tandem repeat analysis. A total of 10,000 viable cells were plated in 96-well microtiter plates in 200 μL of RPMI media with different concentrations of PLX4720, cobimetinib, or both in triplicate. The 48-hour proliferation was assessed using the Cell

We believe that the data support testing regimens that effectively inhibit ERK signaling as treatment of RAS-mutant leukemias, which account for 15% to 30% of patients with CMML and other myeloid malignancies (11, 16). Experience from the treatment of this patient here suggests that intermittent MEK inhibitor administration may enhance pathway inhibition and improve therapeutic efficacy without incurring increased toxicity compared with continuous administration.

We also used quantitative measurements of tumor-derived DNA in plasma to monitor tumor burden dynamically of both malignancies throughout treatment. The burden of BRAFV600E and NRASG12R DNA in plasma correlated with conventional radiographic and hematologic laboratory parameters and confirmed the advantageous effects of combined RAF–MEK inhibition in this patient. In the future, monitoring of tumor-derived DNA in plasma could obviate the need for frequent radiographic monitoring of melanoma with defined genetic alterations present in the bulk melanoma clone.

This case presents an example of ERK activation in RAS-mutant cells induced by vemurafenib and evidence that combined RAF–MEK inhibition offers the potential to treat RAS-mutant disease arising during RAF inhibitor therapy. The observations from the clinical management of this patient present a rationale for intermittent dosing of RAF and MEK inhibitors in the management of BRAF-mutant melanoma and the therapeutic potential of MEK inhibition in refractory RAS-mutant myeloid leukemias.

METHODS

Cell-Free Quantitative Digital PCR of Plasma BRAFV600E and NRASG12R

A TaqMan assay was designed to amplify the region of interest (BRAFV600E or NRASG12R) and distinguish between the wild-type and mutant target using a pair of competitive fluorophore-labeled probes (available upon request). PCR reaction mixtures (20 μL), containing a limited template dilution, were partitioned into approximately 20,000 1-nL droplets using the Bio-Rad QX200 droplet generator system according to the manufacturer’s specifications. The limited template dilution ensured that the estimated number of templates per partition (λ) was within the dynamic range of the instrument. The Poisson correction factor was applied at the analysis stage to account for the eventuality of multiple template occupancy. Following the endpoint PCR amplification, carried under conducive conditions, the individual droplets were analyzed using the Bio-Rad QX200 Droplet Reader system and the proprietary analysis software QuantaLife. The mutant to wild-type ratio and the 95% confidence interval (CI) were calculated as previously described (17). To determine the limit of detection for the designed assays, we measured the lowest detectable amount of mutant target within a large excess of wild-type genomic DNA (gDNA). The observed number of mutant target copies was in close linear relationship with the expected quantities (R² > 0.99), and a single copy of mutant DNA resolved within 10² copies of wild-type DNA (Supplementary Fig. S3).

Leukemia Cell Line Drug Studies

Cell lines were originally obtained from DSMZ or the American Type Culture Collection, and all cell lines were authenticated by Promega short-tandem repeat analysis. A total of 10,000 viable cells were plated in 96-well microtiter plates in 200 μL of RPMI media with different concentrations of PLX4720, cobimetinib, or both in triplicate. The 48-hour proliferation was assessed using the Cell

one of three treatment arms: dabrafenib monotherapy or dabrafenib with one of two doses of trametinib (1 mg or 2 mg/d). Concomitant treatment with 2 mg daily of MEK inhibitor was associated with increased response rate (76% vs. 54%), increased complete response rate (9% vs. 4%), and prolonged median progression-free survival (12 months vs. 5.8 months; ref. 12). In addition to increasing progression-free survival and response rate, combined MEK and RAF inhibition was associated with a decreased frequency of squamous cell carcinomas compared with monotherapy. Inhibitors of MEK and RAF are traditionally administered on schedules designed to inhibit ERK signaling continuously with the presumption that this is required for effective antitumor activity. There are rationales for intermittent therapy, however. The degree of pathway inhibition with MEK inhibitors given daily is limited by toxicity; higher doses and greater pathway inhibition might be achievable with intermittent dosing. Moreover, continuous ERK pathway inhibition causes chronic reactivation of feedback inhibition and other adaptations of the tumor (2, 13). Although the results here come from a single patient, in 2 years of intermittent therapy with RAF inhibition followed by combined RAF–MEK inhibition, no clinical evidence of resistance was apparent. These data suggest that intermittent therapy avoids constant selection for vemurafenib-resistant cells seen with continuous drug administration. In support of this, recent preclinical data with RAF inhibitors suggest that intermittent dosing schedules can delay resistance (13).

The experience of this patient strongly supports testing the efficacy of inhibitors of ERK signaling on intermittent schedules in both preclinical and clinical models. The ability to increase dosage and pathway inhibition while reducing feedback could increase the efficacy of these regimens in tumors with mutant RAS and mutant RAF.

It is worth noting that in this patient, the leukemia was undiagnosed until the patient was treated with the RAF inhibitor. This suggests that the NRAS mutation alone did not elevate ERK output to sufficient levels to cause clinically overt leukemia. It is very likely that the RAF inhibitor enhanced tumor growth by activating ERK signaling. Suppressing ERK activity, by either discontinuing the RAF inhibitor or treating with nontoxic doses of a MEK inhibitor, was sufficient to suppress leukemia growth. Although the patient did not achieve complete remission of the CMML with MEK inhibition, it is possible that the concomitant vemurafenib administration may have blunted the antileukemic effects of cobimetinib by inducing some activation of ERK in the RAS-mutant leukemia cells (as demonstrated in acute myeloid leukemia cell lines; Fig. 2C).

As described earlier, in vitro drug testing suggested that combined RAF and MEK inhibition limited the efficacy of MEK inhibitor to suppress ERK signaling. In this case, it was not possible to give the MEK inhibitor without vemurafenib because of the necessity to treat both tumors. However, two clinical studies of MEK inhibition in myeloid malignancies have been reported (14, 15). An ongoing phase I/II evaluation of trametinib has noted clinical activity in patients with RAS-mutant myeloid malignancies using the recommended phase II dosing of 2 mg daily (14). An overall response rate of 28% of patients with RAS-mutant leukemia has been observed with 11 of 57 RAS-mutant patients experiencing a marrow complete remission.

We believe that the data support testing regimens that effectively inhibit ERK signaling as treatment of RAS-mutant leukemias, which account for 15% to 30% of patients with CMML and other myeloid malignancies (11, 16). Experience from the treatment of this patient here suggests that intermittent MEK inhibitor administration may enhance pathway inhibition and improve therapeutic efficacy without incurring increased toxicity compared with continuous administration.

We also used quantitative measurements of tumor-derived DNA in plasma to monitor tumor burden dynamically of both malignancies throughout treatment. The burden of BRAFV600E and NRASG12R DNA in plasma correlated with conventional radiographic and hematologic laboratory parameters and confirmed the advantageous effects of combined RAF–MEK inhibition in this patient. In the future, monitoring of tumor-derived DNA in plasma could obviate the need for frequent radiographic monitoring of melanoma with defined genetic alterations present in the bulk melanoma clone.

This case presents an example of ERK activation in RAS-mutant cells induced by vemurafenib and evidence that combined RAF–MEK inhibition offers the potential to treat RAS-mutant disease arising during RAF inhibitor therapy. The observations from the clinical management of this patient present a rationale for intermittent dosing of RAF and MEK inhibitors in the management of BRAF-mutant melanoma and the therapeutic potential of MEK inhibition in refractory RAS-mutant myeloid leukemias.

METHODS

Cell-Free Quantitative Digital PCR of Plasma BRAFV600E and NRASG12R

A TaqMan assay was designed to amplify the region of interest (BRAFV600E or NRASG12R) and distinguish between the wild-type and mutant target using a pair of competitive fluorophore-labeled probes (available upon request). PCR reaction mixtures (20 μL), containing a limited template dilution, were partitioned into approximately 20,000 1-nL droplets using the Bio-Rad QX200 droplet generator system according to the manufacturer’s specifications. The limited template dilution ensured that the estimated number of templates per partition (λ) was within the dynamic range of the instrument. The Poisson correction factor was applied at the analysis stage to account for the eventuality of multiple template occupancy. Following the endpoint PCR amplification, carried under conducive conditions, the individual droplets were analyzed using the Bio-Rad QX200 Droplet Reader system and the proprietary analysis software QuantaLife. The mutant to wild-type ratio and the 95% confidence interval (CI) were calculated as previously described (17). To determine the limit of detection for the designed assays, we measured the lowest detectable amount of mutant target within a large excess of wild-type genomic DNA (gDNA). The observed number of mutant target copies was in close linear relationship with the expected quantities (R² > 0.99), and a single copy of mutant DNA resolved within 10² copies of wild-type DNA (Supplementary Fig. S3).

Leukemia Cell Line Drug Studies

Cell lines were originally obtained from DSMZ or the American Type Culture Collection, and all cell lines were authenticated by Promega short-tandem repeat analysis. A total of 10,000 viable cells were plated in 96-well microtiter plates in 200 μL of RPMI media with different concentrations of PLX4720, cobimetinib, or both in triplicate. The 48-hour proliferation was assessed using the Cell
Viability Luminescent Assay Kit (CellTiter-Glo, Promega). Results were normalized to growth of cells in media containing an equivalent volume of dimethyl sulfoxide (DMSO). The concentration at which 50% inhibition in proliferation occurred was determined using GraphPad Prism 5.0 software. For Western blot analysis of signaling pathways, cell lines were exposed to different concentrations of PLX4720, cobimetinib, or both for 4 hours. Cells were then collected and lysed in lysis buffer and separated by electrophoresis. Nitrocellulose membrane was blocked in TBST/5% milk and incubated with antibodies.

**Flow Cytometric Analysis of pERK in Primary Peripheral Blood Mononuclear Cells**

Cryopreserved peripheral blood mononuclear cells were thawed, washed with PBS, fixed, and stained for surface markers to distinguish myeloid cell populations (CD14+) along with intracellular pERK as previously described (5).

**Genetic Analysis**

We used standard techniques to extract gDNA from the melanoma tumor, leukemic BM MNC, and saliva specimens. Barcoded, massively parallel sequencing libraries were prepared (New England Biolabs, Kapa Biosystems), and exon capture was performed on barcoded pools (Nimblegen SeqCap) according to the manufacturer’s directions. Briefly, we designed and synthesized synthetic DNA probes complementary to the coding sequence of 279 genes known to undergo somatic genomic alterations in cancer (Supplementary Table S1). gDNA libraries were subjected to solution-phase hybrid capture using the DNA probes, followed by massively parallel sequencing on the Illumina HiSeq 2500. We sequenced 100 bases from both ends of library DNA fragments, achieving approximately 15 million purity filtered reads per sample. This yielded target gene haploid coverage of 209-, 429-, 388-, and 271-fold from the melanoma, BM MNC number #1 (pre vemurafenib therapy), BM MNC number #2 (on vemurafenib and cobimetinib), and saliva samples, respectively. Paired reads were aligned to the reference human genome using the Burrows-Wheeler Alignment tool (18) and post-processed using the Genome Analysis Toolkit according to best practices (19). Single-nucleotide variants were called using muTecT (20), and indels were called using SomaticIndelDetector (19).

Because somatic mutations in the leukemia were present at low levels in the saliva sample, we additionally retained mutations and indels if the variant alleles frequency in the tumor was >5 times that in the matched normal. All alterations were manually reviewed using the Integrative Genomics Viewer (21).

**Disclosure of Potential Conflicts of Interest**

Margaret K. Callahan has received a commercial research grant from Bristol-Myers Squibb and is a consultant/advisory board member of GlaxoSmithKline and Bristol-Myers Squibb. N. Rosen has received a commercial research grant from Chugai and is a consultant/ advisory board member of Chugai and AstraZeneca. P.B. Chapman is a consultant/advisory board member of Genentech. No potential conflicts of interest were disclosed by the other authors.

**Authors’ Contributions**

**Conception and design:** O. Abdel-Wahab, V.M. Klimek, R. Rampal, M.K. Callahan, D.B. Solit, P.B. Chapman

**Development of methodology:** A.A. Gaskell, M.K. Callahan, T. Merghouh, D.B. Solit, R.L. Levine

**Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.):** O. Abdel-Wahab, V.M. Klimek, A.A. Gaskell, A. Viale, E. Kim, R. Rampal, J.J. Harding, M.K. Callahan, T. Merghouh, M.F. Berger, P.B. Chapman

**Analysis and interpretation of data (e.g., statistical analysis, bios-statistics, computational analysis):** O. Abdel-Wahab, V.M. Klimek, A.A. Gaskell, D. Cheng, E. Kim, R. Rampal, M. Bluh, J.J. Harding, T. Merghouh, M.F. Berger, N. Rosen, R.L. Levine, P.B. Chapman

**Writing, review, and/or revision of the manuscript:** O. Abdel-Wahab, V.M. Klimek, A.A. Gaskell, R. Rampal, J.J. Harding, M.K. Callahan, D.B. Solit, N. Rosen, R.L. Levine, P.B. Chapman

**Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases):** J.J. Harding, M.F. Berger

**Study supervision:** O. Abdel-Wahab, P.B. Chapman

**Acknowledgments**

The authors thank Drs. Sarang Abhyankar and Mika Sovak at Genentech for supplying cobimetinib and, along with Dr. Alice Chung, for helpful suggestions on the article.

**Grant Support**

O. Abdel-Wahab is supported by an NIH K08 Clinical Investigator Award (1K08CA160647-01) and the Josie Robertson Investigator Program. P.B. Chapman is supported in part by the John F. Figge Research Fund.

Received December 31, 2013; revised February 1, 2014; accepted February 26, 2014; published OnlineFirst March 3, 2014.

**REFERENCES**

Combined RAF–MEK Inhibition in RAS- and BRAF-Mutant Malignancies


Efficacy of Intermittent Combined RAF and MEK Inhibition in a Patient with Concurrent BRAF- and NRAS-Mutant Malignancies

Omar Abdel-Wahab, Virginia M. Klimek, Alisa A. Gaskell, et al.

Cancer Discovery 2014;4:538-545. Published OnlineFirst March 3, 2014.

Updated version
Access the most recent version of this article at:
doi:10.1158/2159-8290.CD-13-1038

Supplementary Material
Access the most recent supplemental material at:
http://cancerdiscovery.aacrjournals.org/content/suppl/2014/03/04/2159-8290.CD-13-1038.DC1
http://cancerdiscovery.aacrjournals.org/content/suppl/2021/03/04/2159-8290.CD-13-1038.DC2

Cited articles
This article cites 19 articles, 6 of which you can access for free at:
http://cancerdiscovery.aacrjournals.org/content/4/5/538.full#ref-list-1

Citing articles
This article has been cited by 14 HighWire-hosted articles. Access the articles at:
http://cancerdiscovery.aacrjournals.org/content/4/5/538.full#related-urls

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, use this link
http://cancerdiscovery.aacrjournals.org/content/4/5/538.
Click on "Request Permissions" which will take you to the Copyright Clearance Center's (CCC) Rightslink site.