A First-in-Human Study and Biomarker Analysis of NKTR-214, a Novel IL2Rβγ-Biased Cytokine, in Patients with Advanced or Metastatic Solid Tumors

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ABSTRACT

NKTR-214 (bempegaldesleukin) is a novel IL2 pathway agonist, designed to provide sustained signaling through heterodimeric IL2 receptor βγ to drive increased proliferation and activation of CD8+ T and natural killer cells without unwanted expansion of T regulatory cells (Treg) in the tumor microenvironment. In this first-in-human multicenter phase I study, NKTR-214 administered as an outpatient regimen was well tolerated and showed clinical activity including tumor shrinkage and durable disease stabilization in heavily pretreated patients. Immune activation and increased numbers of immune cells were observed in the periphery across all doses and cycles with no loss of NKTR-214 activity with repeated administration. On-treatment tumor biopsies demonstrated that NKTR-214 promoted immune cell increase with limited increase of Tregs. Transcriptional analysis of tumor biopsies showed that NKTR-214 engaged the IL2 receptor pathway and significantly increased genes associated with an effector phenotype. Based on safety and pharmacodynamic markers, the recommended phase II dose was determined to be 0.006 mg/kg every three weeks.

SIGNIFICANCE: We believe that IL2- and IL2 pathway–targeted agents such as NKTR-214 are key components to an optimal immunotherapy treatment algorithm. Based on its biological activity and tolerability, NKTR-214 is being studied with approved immuno-oncology agents including checkpoint inhibitors. See related commentary by Sullivan, p. 694.

INTRODUCTION

In the treatment of patients with cancer, low levels of tumor-infiltrating lymphocytes (TIL) prior to the introduction of checkpoint inhibitors is predictive of a poor response (1–3). Stimulation of the IL2 receptor (IL2R) pathway is a potent means of expanding and activating immune cells, specifically lymphocytes including cytotoxic CD8+ T cells (4). FDA-approved high-dose (HD) IL2 (aldesleukin) treatment mediates durable clinical benefit in a limited number of patients with melanoma and renal cell carcinoma (RCC), presumably through CD8+ T-cell expansion (5). However, the...
frequent and rapid induction of the IL2 cytokine signaling cascade is associated with an increased risk for serious adverse events (SAE; refs. 6–10). The ability of IL2 to broadly expand many lymphocyte cell populations is a critical feature of its mechanism of action as an immunotherapy and a nonredundant role of IL2 biology.

NKTR-214 (bempedalnesleukin) is a human recombinant IL2 attached to an average of six releasable polyethylene glycol (PEG) chains (11, 12). In the 1980s, PEG-modified IL2 (PEG-IL2) was developed in an effort to improve the solubility and decrease the plasma clearance of IL2 (13). Results of a phase I study describing the use of PEG-IL2 in combination with HD IL2 to treat patients with metastatic RCC and melanoma were reported in 1995 (14). Results from the phase I study concluded that the toxicities and tumor response rates to combination treatment with HD IL2 followed by administration of PEG-IL2 were similar to those of HD unconjugated IL2 alone. Further development of PEG-IL2 was discontinued. In the case of NKTR-214, attachment of PEG chains can also be creatively utilized to not only alter a molecule’s pharmacokinetics but also alter its receptor binding. NKTR-214 is engineered to bias receptor selectivity and alter pharmacokinetics of IL2 (11, 12), while leaving the well-understood amino acid sequence of IL2 (aldesleukin) unaltered. When fully PEGylated, NKTR-214 is a prodrug that has essentially no biological activity. Upon i.v. administration, the PEG chains slowly release to generate active cytokine species with limited binding to the IL2Rα subunit, thereby biasing signaling to favor the dimeric IL2Rαβγ (CD122). Consequently, NKTR-214 increases the proliferation, activation, and effector function of CD8+ T cells and natural killer (NK) cells within the tumor microenvironment (TME) without expanding the undesirable intratumoral T regulatory cells (Treg; ref. 11). In murine tumor models, NKTR-214 generates a ratio of CD8+ cytotoxic T cells to Tregs of >400:1 in tumor, and when combined with an anti-CTLA4 checkpoint inhibitor antibody, there was a strong synergistic effect (11). Based on supportive preclinical efficacy, immunobiology data, and pharmacology and toxicology studies performed in rats and monkeys, a first-in-human study was initiated using NKTR-214 in patients with advanced solid tumors. We report here on the safety, pharmacodynamics, and immunologic and antitumor activity of NKTR-214 monotherapy in patients treated with multiple prior lines of therapy.

RESULTS
Patient Characteristics and Disposition
Twenty-eight patients with advanced or locally advanced solid tumor malignancies were enrolled across three centers from December 2015 to April 2017. Demographics and disease characteristics are summarized in Supplementary Table S1. The majority of patients had a diagnosis of metastatic RCC (n = 15; 53.6%) or melanoma (n = 7; 25.0%). All patients had received prior anticancer treatment: 16 (57.1%) had received targeted therapy; 16 (57.1%) had received an immune checkpoint inhibitor; and 6 (21.4%) had received an immune checkpoint inhibitor in addition to other immuno-

therapy. The median number of prior therapies was 2 (range, 1–12; Supplementary Fig. S1A). The most common reason for study drug discontinuation was progressive disease by RECIST (n = 15; 53.6%) or physician decision (n = 6; 21.4%), primarily for patients who did not respond rapidly and had other therapeutic options. One patient discontinued treatment due to an infusion-related reaction. Patients received a median of 3 infusions of NKTR-214 (range, 1–25 infusions), with a median duration of exposure of 69.5 days (range, 15–533 days).

Safety and Tolerability
All 28 patients enrolled in the study had treatment-emergent adverse events (TEAE), with 26 patients having TEAEs that were considered related to the study drug. TEAEs and treatment-related adverse events (TRAE) occurring in ≥3 patients across all doses are listed in Table 1 and in Supplementary Table S2, respectively. Six of 28 (21.4%) patients reported grade 3 TRAEs; there were no grade 4 or 5 TRAEs. At the highest NKTR-214 dose tested (0.012 mg/kg), 1 patient experienced two dose-limiting toxicities (DLT; grade 3 hypotension and grade 3 syncope), which resolved within 24 hours following i.v. fluid treatment and one dose of tocilizumab. This patient was discharged from the hospital 2 days after admission for the events and continued on study, receiving two additional doses of NKTR-214 0.006 mg/kg that were well tolerated. No other patients were dosed at 0.012 mg/kg. At a NKTR-214 dose of 0.009 mg/kg, 1 patient discontinued the study due to an SAE of grade 3 infusion-related reaction. The reaction occurred following the first dose of NKTR-214 and resolved with treatment. This patient had a history of an infusion-related reaction to a previous immuno-oncology agent. There was one reported immune-related adverse event (irAE) of hypothyroidism associated with NKTR-214, which was treated with replacement therapy. The MTD was defined as the highest dose of NKTR-214 that was administered without causing any unacceptable side effects or AEs. DLTs and AEs were considered related to the study drug.

The most common TRAEs included fatigue (71%), flu-like symptoms (68%), pruritus (64%), hypotension (57%), rash (50%), decreased appetite (46%), and arthralgia and cough (each 32%). Flu-like symptoms were managed with acetaminophen and NSAIDs. The median duration of flu-like symptoms, including fever, was 72 hours and became less pronounced after cycle 2. Rash and pruritus were either self-limited or treated with antihistamines. Hypotension usually occurred around 48 to 72 hours following the first NKTR-214 infusion. Declines in blood pressure following administration of NKTR-214 were generally clinically asymptomatic.

All grade 3 TEAEs of hypotension [5 of 28 (18%) patients] were rapidly reversed with i.v. fluid administration, and no patients discontinued treatment due to hypotension. Risk minimization guidelines, which included oral and i.v. fluids during the first week of dosing, were implemented and successfully reduced the incidence of grade 3 hypotension. The frequency of drug administration, every two weeks (q2w) or q3w at a NKTR-214 dose of 0.006 mg/kg, did not appear to affect the safety profile. Based on safety and pharmacodynamic
### Table 1. TEAEs occurring in 3 or more patients across all doses

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<th>0.012 mg/kg (q3w) (n = 6)</th>
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(continued)
markers, the recommended phase II dose (RP2D) was determined to be 0.006 mg/kg q3w.

Pharmacokinetics and Pharmacodynamics

NKTR-214–related cytokine (NKTR-214-RC) represents a mixture of compounds containing IL2 independent of the PEG-conjugation status, and NKTR-214–active cytokine (NKTR-214-AC) represents a mixture of 2-PEG-IL2 and 1-PEG-IL2, and potentially unconjugated IL2, although this species is often undetectable in patient samples; the mean observed plasma concentration–time profiles are shown in Supplementary Fig. S2A. NKTR-214-RC maximal concentrations were achieved shortly after the end of the infusion and declined monoexponentially thereafter. NKTR-214-RC concentrations increased gradually after dosing with either the q3w or q2w administration schedules a dose cohort and cycles, without indication of accumulating with either the q3w or q2w administration schedules (data not shown). Unconjugated IL2 levels were detected in 76 of 425 samples with reportable results. Unconjugated IL2 concentrations in samples with detectable levels ranged between 0.509 and 4.44 ng/mL with one concentration at 19.4 ng/mL. Out of 425 samples analyzed, 378 samples (89%) had concentrations of unconjugated IL2 that were below the limit of quantitation or less than 1 ng/mL (the lower limit of quantitation was 0.5 ng/mL; data not shown). Soluble CD25 (sCD25) and lymphocyte levels were measured as pharmacodynamic indicators of immune system activation (Supplementary Fig. S2B and S2C). Changes in absolute lymphocyte counts were observed with each q3w treatment cycle and included lymphopenia on day 3 followed by lymphocytosis, which peaked between days 8 and 11, and returned to near-baseline levels prior to the next cycle. Similarly, sCD25 levels increased within each treatment cycle, peaked around day 8, and returned to near-baseline levels prior to each q3w cycle. PD kinetics were observed at all dose levels and at the q2w dosing schedule (data not shown). Anti–NKTR-214 or anti-IL2 antidrug antibodies were not present in any samples tested for this study (data not shown).

Antitumor Activity

Twenty-eight patients were enrolled in the study, and 26 were evaluable for efficacy. Treatment exposure is shown in Supplementary Fig. S1B. Although no objective responses were observed using RECIST criteria, 9 of 26 patients (35%) experienced maximum tumor reductions ranging from 2% to 30%. One patient with RCC had a 40% reduction on the right adrenal gland at the first on-treatment scan [the overall response was stable disease (SD)] and experienced a 30% reduction of the sum of 2 target lesions at the second post-treatment scan; a CT scan of a lung lesion is shown in Supplementary Fig. S1C. Best overall response included one partial response in 14 patients (53.8%; Supplementary Table S3). One patient with RCC had a 40% reduction on the right adrenal gland at the first on-treatment scan [the overall response was stable disease (SD)] and experienced a 30% reduction of the sum of 2 target lesions at the second post-treatment scan; a CT scan of a lung lesion is shown in Supplementary Fig. S1C. Best overall response included one partial response in 14 patients (53.8%; Supplementary Table S3). One patient with metastatic melanoma, previously treated with ipilimumab and a BRAF inhibitor, received 25 cycles of NKTR-214 and had durable SD for 15 months. A second patient with metastatic RCC, who had progressed on HD IL2 and was refractory to single-agent anti-OX40 and nivolumab, was treated with 19 cycles of NKTR-214 and had durable SD for 13 months. Of note, there were 3 immunotherapy-naïve patients with RCC that showed immunologic changes in the blood and tumor, who received standard-of-care nivolumab within 1 month immediately following end of treatment with NKTR-214, and all 3 patients experienced a partial response
NKTR-214 in Patients with Advanced Solid Tumors

at their first post-treatment scan after follow-on therapy with nivolumab.

**Treatment-Related Changes in Peripheral Blood**

To determine if NKTR-214 could induce functional immune cells in the blood, a multiparametric flow cytometry analysis was performed on peripheral blood mononuclear cells (PBMC) collected during the first 2 cycles, before treatment on day 1 and after treatment on day 8. NKTR-214 treatment significantly increased proliferation of CD4+ T cells, CD8+ T cells, and NK cells at day 8 after each cycle (Fig. 1A). Proliferating (Ki-67+ cells) expressed higher levels of inducible costimulator (ICOS) and PD-1 activation markers, compared with nonproliferating (Ki-67−) cells, consistent with an activated phenotype (Fig. 1B). Analysis of the frequencies of CD4+ T cells, CD8+ T cells, and NK cells within the CD45+ immune cell population and absolute number of cells is shown in Supplementary Fig. S3A–S3C. Although there was no increase in the frequencies of peripheral CD4+ and CD8+ T cells within CD45+ immune cells (Supplementary Fig. S3A), treatment with NKTR-214 resulted in a significant increase in the absolute numbers of CD4+ and CD8+ T cells (Supplementary Fig. S3B). There were also increases in the frequencies of CD4+ T cells, CD8+ T cells, and NK cells expressing ICOS and PD-1; increases were observed regardless of dose level or regimen (Supplementary Fig. S4A–S4C).

Expression of costimulatory molecule OX-40, and inhibitory molecule CTLA4, was significantly increased in proliferating NK and T cells, and this appeared to be more pronounced in CD4+ T cells (Supplementary Fig. S3D). An increase in the frequency and absolute number of NK cells after treatment was also noted (Supplementary Fig. S3C). The gating strategy for flow cytometry analysis is shown in Supplementary Fig. S5A and S5B.

**Treatment-Related Changes in the Tumor**

To demonstrate NKTR-214 activated genes associated with the IL2 pathway, targeted analysis of immune networks using NanoString gene analysis to quantify transcript levels was performed on 12 matched baseline and on-treatment tumor biopsies. The Human PanCancer Immune Profiling panel was used to analyze genes associated with immune networks, making it applicable to assess the TME from tumors with diverse pathology. Among the 770 genes assayed, 62 genes were significantly increased in the tumor biopsies on-treatment compared with baseline biopsies (Fig. 2A; Supplementary Table S4), and there were no genes of the 770 genes that were significantly decreased. The top 29 most statistically significant differentially expressed genes with P values <0.01 are listed in Supplementary Table S5. The capacity...
of NKTR-214 to induce immune activation, cytotoxic and effector gene signatures in the tumor was assessed. Our analysis revealed an increased expression of genes associated with T-cell infiltration and signaling (CD3G, CD3D, CD3E, CD247, and ZAP70; P ≤ 0.05), T-cell activation and co-inhibitory molecules (ICOS, TNFRSF9, PDCD1, CTLA4, TIGIT, and LAG3; P ≤ 0.05), and of cytotoxic effector genes (PRF1, GZMB, GZMA, and GZMK; P ≤ 0.05). Genes encoding for PD-L1 and PD-L2 (PDCD1LG2) were also significantly increased, whereas HAVCR2 (TIM3) and MHC class I molecules had no change (Supplementary Fig. S6A). Notably, CD8 and CD4+ Th1-associated genes were increased after treatment (CD8A, CD8B, EOMES, TBX21, and IFNG; P ≤ 0.05), with no significant change noted in Th2, Th17, or inhibitory cytokine-associated genes (IL4, GATA3, IL17A, RORC, TGFB1, TGFB2, and IL10; Fig. 2F and G). Several immune NK-cell genes (KLRK1, KLRD1, KLRK1, and KLRC1; P ≤ 0.05) were increased (Fig. 2H). CXCR3, encoding a chemokine receptor expressed by NK cells, CD4+ Th1, and CD8+ cytotoxic T lymphocytes, is essential for their localization to the tumor and was also increased after treatment (Fig. 2G). Consistently, the gene signatures associated with the presence and activity of T cells including CD4+ Th1 cells, CD8+ T cells, NK cells, and cytotoxic cells were all significantly increased (Supplementary Fig. S6B). Taken together, our results show that NKTR-214 induced a cytotoxic and effector gene signature in the TME consistent with its mechanism of action.

To determine if NKTR-214 promoted an increase in TILs, 14 matched pretreatment and on-treatment tumor biopsies were analyzed by flow cytometry and showed a marked increase in the frequency of CD45R+ live cells in 8 of 14 patients after treatment (Fig. 3A). Similarly, the frequency of NK and CD8+ T cells within live CD45R+ immune cells increased after treatment in 11 of 14 and 9 of 14 patients, respectively (Fig. 3B). To further assess the increase in immune cells within the tumor, the density of infiltrating CD8+ T-cell infiltrate was measured by IHC; the majority of patients, 6 of 10, showed an increase in the number of CD8+ staining per area (mm²) after treatment compared with baseline, and data from 1 patient are shown (Fig. 3C). Because tumor tissue was limited in several patients, not all samples could be analyzed
for flow cytometry, gene expression, and IHC. For example, some patients had samples available for flow cytometry and NanoString analysis but not IHC. However, there were 6 patients for whom both IHC and NanoString data were available (Supplementary Fig. S7A), and there was a good correlation between CD8+ T-cell changes measured by these two different approaches (Supplementary Fig. S7B). In pretreatment and on-treatment tumor biopsies, sequencing of the T-cell receptor β (TCRβ) repertoire showed an increase in T-cell infiltrates in 10 of 12 patients and changes in clonality with a trend toward increased frequency of specific clones after treatment in 8 of 12 patients (Fig. 3D). One patient with melanoma who had received 12 prior lines of therapy was analyzed for changes in productive clonality and clonal frequency, and NKTR-214 appeared to induce an increase in both existing and new clones at week 3 (Supplementary Fig. S8A). Furthermore, an increase in clonality was associated with increased T-cell infiltration in 7 of 12 patients, reflecting the induction of a more oligoclonal T-cell repertoire (Fig. 3D; Supplementary Fig. S8B).

Despite the robust proliferation and increase in the absolute number of CD4+CD25hiFOXP3+ Tregs in the peripheral blood after treatment (Fig. 3E; Supplementary Fig. S9A and S9B) and upregulation of markers associated with enhanced suppressive function (ICOS and CTLA4; Supplementary Fig. S9C; refs. 4, 15), NKTR-214 administration did not appear to have the same effect on CD4+CD25hiFOXP3+ Tregs in the TME as in the peripheral blood. Flow cytometry analysis showed that the percentages of CD4+CD25hiFOXP3+ Tregs were slightly increased in 4 patients and either reduced or did not change in 10 patients (Fig. 3F). From the same tumor biopsies analyzed, FOXP3 staining by IHC showed little to no increase in FOXP3+ cells within total CD4+ T cells or CD8+ T cells (bottom plot) in tumor biopsy samples analyzed by flow cytometry, n = 14. G, Quantification of the number of FOXP3+ cells by IHC, n = 10, and each circle represents one individual tumor sample with red circles indicating increased and black circles indicating unchanged or reduced FOXP3+ cells on-treatment compared with pretreatment. H, PD-1 expression within CD4+ (top plot) and CD8+ T cells (bottom plot) in tumor biopsy samples was analyzed by flow cytometry (n = 14).

Figure 3. Intratumoral changes in critical lymphocyte populations and numbers after NKTR-214 treatment. Tumor biopsies were obtained at baseline (blue) and after NKTR-214 treatment (orange). Immune infiltrates, including CD45+ live cells, NK cells, and CD8+ T cells, were analyzed by flow cytometry on freshly disaggregated tumor tissue. Patients #13 and #14 received a NKTR-214 dose of 0.006 mg/kg q2w. A, Percentage of live CD45+ cells within total cells. B, Percentage of NK cells (top plot) and CD8+ T cells (bottom plot) within CD45+ live cells. C, A portion of the tumor biopsy from the chest wall of a patient with RCC (13) dosed q2w was further analyzed for CD8 staining by IHC (left). 10 patients had pretreatment and on-treatment IHC data (right), and each circle represents one individual tumor sample with red circles indicating increased and black circles indicating similar or reduced number of CD8+ T cells on-treatment compared with pretreatment. D, TCR sequencing examining the percentage of T-cell infiltration (the fraction productive of cells mass estimate values obtained through the immuSeq Analyzer software; top plot) and clonality (bottom plot) in tumor biopsy samples, n = 12. E, Percentage of CD4+CD25hiFOXP3+ Tregs within total CD4+ T cells in the blood before and 8 days after treatment with NKTR-214. A representative result is shown on the left. Paired t test, **, P < 0.01. F, Percentage of CD4+CD25hiFOXP3+ Tregs within CD8+ T cells in tumor biopsy samples was analyzed by flow cytometry, n = 14. G, Quantification of the number of FOXP3+ cells by IHC, n = 10, and each circle represents one individual tumor sample with red circles indicating increased and black circles indicating unchanged or reduced FOXP3+ cells on-treatment compared with pretreatment. H, PD-1 expression within CD4+ (top plot) and CD8+ T cells (bottom plot) in tumor biopsy samples was analyzed by flow cytometry (n = 14).
DISCUSSION

This is a first-in-human report of NKTR-214. Treatment with NKTR-214 was well tolerated and feasible as an outpatient regimen. Grade 3 TRAEs were experienced by 21.4% of patients; there were no grade 4 TRAEs or any treatment-related deaths. The most common TRAEs were of grade 1 and 2 severity and included flu-like symptoms, rash, and pruritus, the majority of which coincided with the peak plasma concentrations of the active cytokine and resolved spontaneously or were mitigated by nonprescription oral or topical treatments. NKTR-214–related hypotension was predictable, manageable, and reversible. The incidence of grade 3 hypotension was reduced once hypotension risk mitigation strategies were implemented. Notably, 3 of the 5 patients who experienced grade 3 hypotension had preexisting adrenal insufficiency, were managed conservatively, and continued to be dosed on the study. The Safety Review Committee (SRC) decided to halt enrollment into the 0.012 mg/kg cohort after dosing 1 patient. This decision was based on the composite of hypotension, syncpe, and cytokine release syndrome that necessitated a hospital admission of this patient and the goal to identify a NKTR-214 dose that could be delivered as a safe, outpatient treatment. Based on the tolerability, convenient dosing schedule, and pharmacodynamic changes at all NKTR-214 doses tested, a NKTR-214 dose of 0.006 mg/kg q3w was selected as the RP2D.

NKTR-214 engaged the IL2R pathway and provided robust systemic immune activation with repeated administration, as demonstrated by dynamic changes in absolute lymphocyte counts and sCD25. Most importantly, these pharmacodynamic changes did not wane after repeated cycles. NKTR-214 induced robust proliferation of CD4+ T cells, CD8+ T cells, and NK cells in the blood, and the cells were activated as indicated by high levels of ICOS, PD-1, CTLA4, and OX40 expression. Importantly, treatment with NKTR-214 resulted in a significant increase in the absolute numbers of CD4+ T cells, CD8+ T cells, and NK cells. The induction of CD4+ T-cell, CD8+ T-cell, and NK-cell proliferation in the peripheral blood was also observed in patients treated with human recombinant IL15 (rIL15), and these changes were most evident in NK and CD8+ T cells and to a lesser degree in CD4+ T cells (16). IL15 shares the IL2/IL15Rα/T-cell, CD8+ T-cell, and NK-cell proliferation and does not cause substantial toxicity necessitating intense supportive care. However, the use of rIL15 as a therapeutic agent may not be optimal due to the low expression of IL15Rβ and the need for high doses to achieve biological responses in vivo (17). The administration of rIL15 as a daily i.v. bolus infusion for 12 consecutive days to 5 patients with metastatic malignant melanoma or metastatic RCC proved to be difficult due to clinical toxicities produced by intense cytokine secretion that occurred in the first 2 hours after treatment (16).

Importantly, treatment with NKTR-214 induced meaningful immunologic changes within the TME. At the RP2D, freshly isolated TILs showed that there was a 15.1-, 3.5-, and 0.7-fold increase in CD8+ T cells, NK cells, and Treg cells, respectively (Supplementary Fig. S10). Transcriptional analysis of the on-treatment tumor biopsies highlighted differentially expressed genes that were significantly increased on-treatment, which support a fundamental reshaping of the TME toward immune cell activation and effector functions. NKTR-214 also triggered immune regulatory pathways as seen in the increased gene expression of PD-L1/PD-L2, TIGIT, LAG3, SOCI, and IDO1. Interestingly, NKTR-214 induced a gene signature associated with the presence and activity of T cells including Th1 cells and CD8+ T cells. The increase of this Th1/CD8+ gene signature was not associated with an increase in the gene signature representing Th2 or Th17 cells that are known to be mediators of inflammation associated with anti–PD-1 and anti-CTLA4 therapy–induced irAEs (18, 19). A study of patients with melanoma treated with anti-CTLA4 combined with HD IL2 demonstrated a trend toward a decreased incidence of grade 3/4 irAEs compared with patients receiving anti-CTLA4 without IL2 (17% vs. 29% and 32%; refs. 20, 21). Prieto and colleagues proposed an elegant theory that the Tregs induced by HD IL2 may decrease the activity of autoreactive T cells to reduce the incidence of irAEs (21). It is well established that IL2 signals control the polarization of CD4+ T-cell subsets leading to enhancement of Th1 and Tregs while limiting the differentiation into Th17 (22, 23). Several mechanisms have been proposed to explain the negative effect of IL2 on Th17, including lowering the expression of the IL6Rβ on CD4+ T cells, thereby reducing the IL6-mediated STAT3 phosphorylation that is essential for the induction of Th17 (24). Other studies showed that the IL2 signaling through STAT3, which competes with STAT5 for binding the IL17 gene locus, constrains the generation of Th17 (25, 26). Hence, the immune effect of NKTR-214 on the tumor appeared to be Th1-dominant with no induction of Th2 or Th17 gene signatures. Despite the increases in Tregs in the periphery with NKTR-214 administration, expansion of these cells was limited in the tumor. Preclinical evidence from mouse tumor models indicates that NKTR-214 limits intratumoral Treg proliferation and survival by way of promoting apoptosis, thereby polarizing the TME toward CD8+ T cells and consequently driving a very high CD8+/Treg ratio (12, 27). Exploratory studies are in progress to further probe the mechanisms by which peripheral Tregs are preserved while intratumoral Tregs are not.

A key immunologic observation was the increase in the percentage of proliferating CD8+ T cells also expressing increased levels of PD-1 in the TME (Fig. 3H). Studies have demonstrated that PD-1 identifies CD8+ tumor-reactive T cells in infiltrating tumors (2, 28–31). The increase in the percentage of CD8+PD-1+ T cells in patients’ tumors and the tumor shrinkage observed in 35% of patients, including one unconfirmed partial response, is promising. Durable SD greater than 1 year was seen in 2 patients, with 1 patient having failed 3 prior immunotherapies. It is possible that compensatory mechanisms, namely PD-L1/PD-L2 (Fig. 2E), may have dampened the potential of these CD8+PD-1+ T cells to convert patients into responders (31–33). Intriguingly, 3 patients with RCC who were anti–PD-1 treatment-naïve experienced TIL elevation with tumor reductions of 1%, 10%, and 20%; all 3 patients ended NKTR-214 with SD and within...
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1 month started anti-PD-1 treatment and experienced rapid tumor reductions at first scan on-treatment, resulting in partial responses (34). These anecdotal observations supported the hypothesis that NKTR-214 treatment may have conditioned the TME by expanding activated TILs, thereby potentially providing synergy with therapies that block inhibitory signals, such as those of the PD-1/PD-L1 axis.

In summary, every patient evaluated showed evidence for activation of the immune response, at all four doses tested, and these effects were reproduced with repeated administration. Given the small number of patients evaluable at q2w (n = 4), no conclusions can be made on the impact of immune activation with a shorter frequency of dosing. Based on the biological activity and tolerability of NKTR-214 as well as the nonoverlapping toxicities with approved checkpoint inhibitors, NKTR-214 is being combined with approved anti-PD-1/PD-L1/CTLA4 agents in ongoing clinical trials. NKTR-214, as a T-cell and NK-cell growth factor, may serve as a central mechanism for boosting tumor-specific T-cell proliferation and activation in the TME and has the potential to serve as a backbone therapy for combination therapy with a variety of agents.

METHODS

Patients and Study Design

This was a multicenter, open-label dose-escalation study using a 3+3 design. Eligible patients were treatment-refractory to ≥1 prior therapy for advanced or locally advanced disease. Twenty-eight patients with a histologically confirmed diagnosis of RCC, melanoma, urachal carcinoma, breast cancer, leiomyosarcoma, chondrosarcoma, or colorectal cancer not amenable to curative therapy were enrolled. All patients provided written informed consent before any protocol-specified procedures. The primary objectives were to assess the safety, MTD, RP2D, and antitumor clinical activity. Secondary objectives were to characterize pharmacokinetics, pharmacodynamics, and immunologic changes in blood and tumor. NKTR-214 was administered at 0.003, 0.006, 0.009, and 0.012 mg/kg as a 15-minute i.v. infusion q3w or 0.006 mg/kg q2w until disease progression or unacceptable toxicities occurred. Patients were held for 24 hours after an infusion center after the first dose of NKTR-214; however, subsequent cycles were administered on an outpatient basis. If no DLTs were observed in a cohort after 21 days, the dose was escalated for the following cohort after review by the SRC, which consisted of the sponsor and at least one site investigator. Additional patients were added to a lower-dose cohort to further assess the safety and tolerability. All patients who received study treatment and one post-treatment scan were evaluable for antitumor activity. Investigator-assessed antitumor activity was evaluated using RECIST 1.1. Baseline tumor imaging was completed within 4 weeks before starting therapy and every 8 weeks thereafter. Tumor responses were confirmed with a repeat scan at least 4 weeks later.

Safety

All patients who received any study drug were included in the safety analyses. AEs, SAEs, and laboratory abnormalities were graded per the National Cancer Institute Common Terminology Criteria for Adverse Events, Version 4.03, and monitored throughout the study until 30 days after the last dose of study drug. TRAEs and TEAEs were reported throughout the observation period and recorded once per patient at the highest grade. DLT was defined as any nontransient grade 3, 4, or 5 hematologic or nonhematologic toxicity that occurred within the first 21 days after dosing and was deemed related or possibly related to the administration of NKTR-214. The MTD was defined as the highest dose of NKTR-214 that was administered without causing any unacceptable side effects or AEs. trAEs were considered AEs of special interest due to their potential clinical significance.

Biomarker Analysis

Serial blood and tumor tissue samples were collected to perform in-depth analysis of immune cells using flow cytometry, nCounter gene expression assay (NanoString), IHC, and TCR sequencing; details are provided in the Supplementary Information.

Statistical Methods

Summary statistics were used for any continuous variables; numbers and percentage were used for categorical variables. For all analyses, baseline values were defined as the last value before the first dose of NKTR-214. All statistical calculations were performed using Statistical Analysis System (SAS) version 9.4 (SAS Institute). Statistical analyses for immune profiling by flow cytometry were performed with GraphPad Prism 7.0 (GraphPad Software). The significance of the difference between groups in the experiments was evaluated by a two-tailed paired t test. A value of P ≤ 0.05 was considered statistically significant.

Disclosure of Potential Conflicts of Interest

M.E. Flavowitz reports receiving commercial research grants from Apexigen, Astellas, Innocrin, Iovance, Merck, Nektar Therapeutics, Novartis, Pfizer, Progenics, Sanofi Aventis, Seattle Genetics, AstraZeneca, Bayer, Bristol-Myers Squibb, Corvus, Eli Lilly, Endocyte, Genentech, and Genmab; is a consultant/advisory board member for Nektar Therapeutics, Janssen, and CRISPR Therapeutics; and reports the receipt of remuneration from Gamida Cell. H.M. Kluger reports receiving commercial research grants from Bristol-Myers Squibb, Merck, and Apexigen; is a consultant/advisory board member for Nektar, Iovance, Merck, Genentech, Corvus, Immunocore, Biodexis, Apexigen, and Alexion; and reports receiving other remuneration from Pfizer and Nektar. M.T. Tetzlaff is a consultant/advisory board member for Novartis LLC, Myriad Genetics, and Seattle Genetics. M.A. Tagliaferri is CMO at Nektar and has ownership interest (including stock, patents, etc.) in the same. J. Zalesky is Chief Scientific Officer at Nektar Therapeutics and has ownership interest (including stock, patents, etc.) in the same. U. Hoch has ownership interest (including stock, patents, etc.) in Nektar. C. Fanton has ownership interest (including stock, patents, etc.) in Nektar Therapeutics. S. Aung has ownership interest (including stock, patents, etc.) in Nektar Therapeutics. P. Hwu is a consultant/advisory board member for Dragonfly, GlaxoSmithKline, Immatics, and Sanofi. N.M. Tannir reports receiving a commercial research grant from Nektar Therapeutics and is a consultant/advisory board member for the same. M. Szolol is a consultant/advisory board member for Genentech-Roche, Bristol-Myers, Merck US, Theravance, Biodexis, Vaccinex, Janssen/Johnson & Johnson, Modulate Therapeutics, Baxalta-Shire, Incyte, Newlink Genetics, Lions Biotechnologies, AstraZeneca/MedImmune, Agonox, Arbutus, Celldex, Inovio, Gritstone, Molecular Partners, Innate Pharma, AbbVie, Immunocore, Genmab, Pfizer, Almac, Hinge, Allakos, Anaoeropharma, Array, Symphogen-Scientific (advisory board), Adaptimmune-Scientific (advisory board), Omnimox-Scientific (advisory board), Lycera (formerly on scientific advisory board), Pieris (scientific advisory board), Novartis, AmphiVena (stock options only), Adaptive Biotechnologies (stock options only), Intensity (stock options only), Argenx (stock options only), Torque, Torque (stock options), Kyowa-Kirin, Seattle Genetics, Nektar, Pierre-Fabre, and Lilly. A. Diab reports receiving a commercial research grant from Nektar Therapeutics and is a consultant/advisory board member for the same. No potential conflicts of interest were disclosed by the other authors.

Statistical Methods for Immune Profiling

The difference between baseline and post-treatment values was evaluated by the Wilcoxon signed-rank test. A value of P ≤ 0.05 was considered statistically significant.

Statistical Analysis for Immune Profiling

The difference between groups in the experiments was evaluated by a two-tailed paired t test. A value of P ≤ 0.05 was considered statistically significant.

Disclosure of Potential Conflicts of Interest

M.E. Flavowitz reports receiving commercial research grants from Apexigen, Astellas, Innocrin, Iovance, Merck, Nektar Therapeutics, Novartis, Pfizer, Progenics, Sanofi Aventis, Seattle Genetics, AstraZeneca, Bayer, Bristol-Myers Squibb, Corvus, Eli Lilly, Endocyte, Genentech, and Genmab; is a consultant/advisory board member for Nektar Therapeutics, Janssen, and CRISPR Therapeutics; and reports the receipt of remuneration from Gamida Cell. H.M. Kluger reports receiving commercial research grants from Bristol-Myers Squibb, Merck, and Apexigen; is a consultant/advisory board member for Nektar, Iovance, Merck, Genentech, Corvus, Immunocore, Biodexis, Apexigen, and Alexion; and reports receiving other remuneration from Pfizer and Nektar. M.T. Tetzlaff is a consultant/advisory board member for Novartis LLC, Myriad Genetics, and Seattle Genetics. M.A. Tagliaferri is CMO at Nektar and has ownership interest (including stock, patents, etc.) in the same. J. Zalesky is Chief Scientific Officer at Nektar Therapeutics and has ownership interest (including stock, patents, etc.) in the same. U. Hoch has ownership interest (including stock, patents, etc.) in Nektar. C. Fanton has ownership interest (including stock, patents, etc.) in Nektar Therapeutics. S. Aung has ownership interest (including stock, patents, etc.) in Nektar Therapeutics. P. Hwu is a consultant/advisory board member for Dragonfly, GlaxoSmithKline, Immatics, and Sanofi. N.M. Tannir reports receiving a commercial research grant from Nektar Therapeutics and is a consultant/advisory board member for the same. M. Szolol is a consultant/advisory board member for Genentech-Roche, Bristol-Myers, Merck US, Theravance, Biodexis, Vaccinex, Janssen/Johnson & Johnson, Modulate Therapeutics, Baxalta-Shire, Incyte, Newlink Genetics, Lions Biotechnologies, AstraZeneca/MedImmune, Agonox, Arbutus, Celldex, Inovio, Gritstone, Molecular Partners, Innate Pharma, AbbVie, Immunocore, Genmab, Pfizer, Almac, Hinge, Allakos, Anaoeropharma, Array, Symphogen-Scientific (advisory board), Adaptimmune-Scientific (advisory board), Omnimox-Scientific (advisory board), Lycera (formerly on scientific advisory board), Pieris (scientific advisory board), Novartis, AmphiVena (stock options only), Adaptive Biotechnologies (stock options only), Intensity (stock options only), Argenx (stock options only), Torque, Torque (stock options), Kyowa-Kirin, Seattle Genetics, Nektar, Pierre-Fabre, and Lilly. A. Diab reports receiving a commercial research grant from Nektar Therapeutics and is a consultant/advisory board member for the same. No potential conflicts of interest were disclosed by the other authors.
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RESEARCH BRIEF

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NKTR-214 in Patients with Advanced Solid Tumors


A First-in-Human Study and Biomarker Analysis of NKTR-214, a Novel IL2R βγ-Biased Cytokine, in Patients with Advanced or Metastatic Solid Tumors

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