RESEARCH ARTICLE

Phase I, Dose-Escalation, Two-Part Trial of the PARP Inhibitor Talazoparib in Patients with Advanced Germline BRCA1/2 Mutations and Selected Sporadic Cancers

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ABSTRACT

Talazoparib inhibits PARP catalytic activity, trapping PARP1 on damaged DNA and causing cell death in BRCA1/2-mutated cells. We evaluated talazoparib therapy in this two-part, phase I, first-in-human trial. Antitumor activity, MTD, pharmacokinetics, and pharmacodynamics of once-daily talazoparib were determined in an open-label, multicenter, dose-escalation study (NCT01286987). The MTD was 1.0 mg/day, with an elimination half-life of 50 hours. Treatment-related adverse events included fatigue (26/71 patients; 37%) and anemia (25/71 patients; 35%). Grade 3 to 4 adverse events included anemia (17/71 patients; 24%) and thrombocytopenia (13/71 patients; 18%). Sustained PARP inhibition was observed at doses ≥0.60 mg/day. At 1.0 mg/day, confirmed responses were observed in 7 of 14 (50%) and 5 of 12 (42%) patients with BRCA mutation-associated breast and ovarian cancers, respectively, and in patients with pancreatic and small cell lung cancer. Talazoparib demonstrated single-agent antitumor activity and was well tolerated in patients at the recommended dose of 1.0 mg/day.

SIGNIFICANCE: In this clinical trial, we show that talazoparib has single-agent antitumor activity and a tolerable safety profile. At its recommended phase II dose of 1.0 mg/day, confirmed responses were observed in patients with BRCA mutation-associated breast and ovarian cancers and in patients with pancreatic and small cell lung cancer. Cancer Discov; 7(6); 1–10. © 2017 AACR.

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L.A. Byers and Z.A. Wainberg are co–senior authors of this article.

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INTRODUCTION

The most-studied PARP enzymes are PARP1 and PARP2, which play critical roles in DNA damage detection and repair (1, 2), including the repair of single-strand DNA breaks through the base excision repair pathway (3–5). It has been hypothesized that single-strand DNA breaks persist when PARP function is compromised, leading to the creation of double-strand DNA breaks during replication (6); these double-strand DNA breaks are usually repaired by homologous recombination repair (HRR), allowing replication to continue (6). However, loss of PARP activity becomes lethal when HRR is compromised. This phenomenon, known as synthetic lethality, is well established for deleterious mutations of \textit{BRCA1} and \textit{BRCA2} (7–9).

The PARP inhibitor olaparib was recently approved for the treatment of advanced ovarian cancer and remains the only approved agent. PARP inhibitors have also demonstrated antitumor activity against other tumor types with DNA repair deficiencies, including breast and prostate cancers (10–13). Talazoparib (also known as MDV3800 and BMN 673) is a novel, potent, and selective inhibitor of PARP1/2 that achieves antitumor cell responses and elicits DNA repair markers at notably lower concentrations than earlier-generation PARP1/2 inhibitors (14, 15). In addition to inhibiting PARP catalytic activity, talazoparib is currently the most potent PARP1/2 inhibitor \textit{in vitro} at trapping PARP–DNA complexes at sites of single-strand DNA breaks (16). Preclinically, talazoparib has favorable metabolic stability, oral bioavailability, and pharmacokinetics (PK) that support its daily schedule in clinical trials (14).

We conducted a first-in-human, phase I dose-escalation (Part 1) trial of talazoparib in patients with advanced solid malignancies and an expansion cohort (Part 2) in patients with tumors predicted to be potentially sensitive to PARP inhibition. These included tumors harboring germline \textit{BRCA1/2} mutations; triple-negative breast cancers; high-grade serous and/or undifferentiated ovarian, fallopian tube, or peritoneal cancers; and castration-resistant prostate and pancreatic cancers. Patients with Ewing sarcoma or small cell lung cancer (SCLC) were also studied; the former was based on a 1,000-cell line screen demonstrating antitumor activity...
(17, 18), and the latter was based on SCLC platinum sensitivity, increased PARP1 expression, and sensitivity of SCLC cell lines and animal models to PARP inhibition (19, 20).

RESULTS

Between January 3, 2011, and August 21, 2014, 113 patients with advanced solid tumors were enrolled at a total of six centers: five in the United States and one in the United Kingdom. A total of 110 patients received talazoparib (Table 1). Thirty-nine patients participated in Part 1 and received talazoparib at nine dose levels ranging from 0.025 to 1.1 mg/day (Fig. 1). An additional 71 patients were treated with talazoparib 1.0 mg/day in Part 2. As of the date of database cutoff (March 31, 2015), 2 patients in Part 1 and 5 patients in Part 2 continue to be treated (Fig. 1).

<table>
<thead>
<tr>
<th>Demographic parameter</th>
<th>Dose escalation (Part 1; n = 39)</th>
<th>Dose expansion (Part 2; n = 71)</th>
<th>Overall (N = 110)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Median age, years (range)</td>
<td>58.0 (19–81)</td>
<td>57.0 (18–88)</td>
<td>57.0 (18–88)</td>
</tr>
<tr>
<td>Male, n (%)</td>
<td>6 (15.4)</td>
<td>28 (39.4)</td>
<td>34 (30.9)</td>
</tr>
<tr>
<td>ECOG performance status, n (%)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0</td>
<td>23 (59.0)</td>
<td>37 (52.1)</td>
<td>60 (54.5)</td>
</tr>
<tr>
<td>1</td>
<td>16 (41.0)</td>
<td>34 (47.9)</td>
<td>50 (45.5)</td>
</tr>
<tr>
<td>Tumor type, n (%)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Breast</td>
<td>8 (20.5)</td>
<td>12 (16.9)</td>
<td>20 (18.2)</td>
</tr>
<tr>
<td>Ovarian/peritoneal</td>
<td>23 (59.0)</td>
<td>11 (15.5)</td>
<td>34 (30.9)</td>
</tr>
<tr>
<td>Prostate</td>
<td>1 (2.6)</td>
<td>3 (4.2)</td>
<td>4 (3.6)</td>
</tr>
<tr>
<td>Pancreatic</td>
<td>3 (7.7)</td>
<td>10 (14.1)</td>
<td>13 (11.8)</td>
</tr>
<tr>
<td>Ewing sarcoma</td>
<td>2 (5.1)</td>
<td>12 (16.9)</td>
<td>14 (12.7)</td>
</tr>
<tr>
<td>SCLC</td>
<td>0</td>
<td>23 (32.4)</td>
<td>23 (20.9)</td>
</tr>
<tr>
<td>Colorectal</td>
<td>2 (5.1)</td>
<td>0</td>
<td>2 (1.8)</td>
</tr>
<tr>
<td>Deleterious mutation, n (%)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>gBRCA1</td>
<td>16 (41.0)</td>
<td>13 (18.3)</td>
<td>29 (26.4)</td>
</tr>
<tr>
<td>gBRCA2</td>
<td>7 (17.9)</td>
<td>20 (28.2)</td>
<td>27 (24.5)</td>
</tr>
<tr>
<td>gBRCA1/2</td>
<td>1 (2.6)</td>
<td>2 (2.8)</td>
<td>3 (2.7)</td>
</tr>
<tr>
<td>Median prior chemotherapy regimens, n (range)</td>
<td>4.0 (1.0–13.0)</td>
<td>2.0 (0.0–6.0)</td>
<td>2.5 (0.0–13.0)</td>
</tr>
<tr>
<td>Median prior platinum regimens, n (range)</td>
<td>2.0 (0.0–4.0)</td>
<td>1.0 (0.0–4.0)</td>
<td>1.0 (0.0–4.0)</td>
</tr>
</tbody>
</table>

Abbreviations: ECOG, Eastern Cooperative Oncology Group; gBRCA, germline BRCA mutated.

**Figure 1.** Patient enrollment and disposition. Abbreviation: ECOG PS, Eastern Cooperative Oncology Group Performance Status.
Safety

The number of patients per dose level, observed dose-limiting toxicities (DLT), dose reductions, and median time on study are provided in Table 2. Dose-limiting thrombocytopenia in cycle 1 occurred in 1 of 6 patients at 0.9 mg/day and 2 of 6 patients assessable for DLT at 1.1 mg/day. The patient treated at 0.9 mg/day experienced grade 3 thrombocytopenia with grade 3 anemia. Of the 2 patients treated at 1.1 mg/day, both experienced grade 3 thrombocytopenia; for 1 of these patients, it became grade 4 thrombocytopenia. All DLTs resolved after temporary interruption of study drug; no hemorrhage was noted. Because 2 patients experienced a DLT at the 1.1 mg/day dose level, an interim dose of 1.0 mg/day was investigated. No DLTs were observed at this dose level in a group of 6 assessable patients. This dose was therefore determined to be the MTD and the recommended dose for Part 2.

In Part 2, 71 patients received talazoparib at 1.0 mg/day via continuous daily dosing. The median relative dose intensity was high at 97.2%, and the dose was well tolerated. Table 2 presents the most common toxicities at this dose related to the study drug, including fatigue (37%), anemia (35%), nausea (32%), thrombocytopenia (21%), alopecia (20%), and neutropenia (15%). Grade 3 or 4 adverse events (AE) assessed by investigator as related were reported in 32 (45%) patients, with the most frequent being anemia (23%), thrombocytopenia (18%), and neutropenia (10%).

Of the 77 patients receiving the 1.0 mg/day dose, 26 patients (34%) reported at least one dose reduction, the majority of whom (20 patients) had reductions due to an AE such as anemia, thrombocytopenia, and neutropenia. Although transient dose holidays were needed as a result of these AEs, no patients permanently withdrew from treatment because of them in either Part 1 or Part 2 of the trial.

There were eight deaths associated with an AE during the study, none of which were considered to be related to study treatment. Two of the deaths occurred in patients with breast cancer enrolled in Part 1 at the entry dose of 1.1 mg/day talazoparib (both related to disease progression). Six of the deaths occurred in patients in Part 2 at the 1.0 mg/day dose of talazoparib (2 patients with pancreatic cancer, both from disease progression; 2 patients with Ewing sarcoma, 1 from dyspnea and the other from respiratory failure; and 2 patients with SCLC, 1 from hypoxia secondary to lung metastases and the other from lung infection).

### Table 2. Part 1 dose escalation schema, DLTs, dose reductions, and common AEs (>15%) or grade 3 to 4 AEs (>4%) assessed by investigator as related at the recommended dose

<table>
<thead>
<tr>
<th>Dose level</th>
<th>Patients (n = 39)</th>
<th>DLTs in first cycle</th>
<th>Dose reductions (any cycle)</th>
<th>Number of treatment days</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Number</td>
<td>Description</td>
<td>Number</td>
<td>Median (range)</td>
</tr>
<tr>
<td>0.025 mg</td>
<td>3</td>
<td>0</td>
<td>2</td>
<td>35 (35–98)</td>
</tr>
<tr>
<td>0.05 mg</td>
<td>3</td>
<td>0</td>
<td>2</td>
<td>99 (34–205)</td>
</tr>
<tr>
<td>0.10 mg</td>
<td>3</td>
<td>0</td>
<td>2</td>
<td>119 (65–253)</td>
</tr>
<tr>
<td>0.20 mg</td>
<td>3</td>
<td>0</td>
<td>2</td>
<td>281 (35–427)</td>
</tr>
<tr>
<td>0.40 mg</td>
<td>3</td>
<td>0</td>
<td>1</td>
<td>226 (97–268)</td>
</tr>
<tr>
<td>0.60 mg</td>
<td>6</td>
<td>0</td>
<td>4</td>
<td>185 (58–289)</td>
</tr>
<tr>
<td>0.90 mg</td>
<td>6</td>
<td>1 Grade 3 TCP</td>
<td>5</td>
<td>261 (30–1114)</td>
</tr>
<tr>
<td>1.00 mg</td>
<td>6</td>
<td>0</td>
<td>5</td>
<td>214 (84–960)</td>
</tr>
<tr>
<td>1.10 mg</td>
<td>6*</td>
<td>2 Grade 3–4 TCP</td>
<td>4</td>
<td>60 (14–196)</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Adverse event</th>
<th>All grade (n = 71)</th>
<th>Grade 3–4 (n = 71)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Any treatment-emergent AE, n (%)</td>
<td>55 (77)</td>
<td>32 (45)</td>
</tr>
<tr>
<td>Blood and lymphatic system disorders, n (%)</td>
<td>40 (56)</td>
<td>30 (42)</td>
</tr>
<tr>
<td>Anemia</td>
<td>25 (35)</td>
<td>16 (23)</td>
</tr>
<tr>
<td>TCP</td>
<td>15 (21)</td>
<td>13 (18)</td>
</tr>
<tr>
<td>Neutropenia</td>
<td>11 (15)</td>
<td>7 (10)</td>
</tr>
<tr>
<td>Gastrointestinal disorders, n (%)</td>
<td>27 (38)</td>
<td>—</td>
</tr>
<tr>
<td>Nausea</td>
<td>23 (32)</td>
<td>—</td>
</tr>
<tr>
<td>General disorders and administration site conditions, n (%)</td>
<td>27 (38)</td>
<td>2 (3)</td>
</tr>
<tr>
<td>Fatigue</td>
<td>26 (37)</td>
<td>2 (3)</td>
</tr>
<tr>
<td>Skin and subcutaneous tissue disorders, n (%)</td>
<td>19 (27)</td>
<td>—</td>
</tr>
<tr>
<td>Alopecia</td>
<td>14 (20)</td>
<td>—</td>
</tr>
</tbody>
</table>

Abbreviation: TCP, thrombocytopenia.

*One patient discontinued from the trial on study day 21 for progressive disease, having received only 8 days of continuous dosing.
Pharmacokinetics

Mean talazoparib plasma concentration–time profiles following single and multiple doses of talazoparib are provided in Fig. 2A–D. Talazoparib PK parameters resulting from the analysis of the plasma concentration–time profiles are provided in Table 3.

Talazoparib demonstrated rapid absorption, with maximum plasma concentration (C_max) generally reached within 2 hours after all evaluated doses and following both single and multiple daily dosing. Steady-state plasma concentrations were reached by 2 weeks of daily dosing across all doses evaluated. Talazoparib was well distributed into tissue compartments, with apparent volume of distribution (Vz/F) estimates in excess of the volume of the systemic circulatory space. Plasma elimination followed biphasic kinetics with a long terminal half-life (t_1/2). Linear elimination across dose levels was apparent following both single and multiple daily dosing as evidenced by parallel terminal phases of the log-linear profiles and similar apparent oral clearance (CL/F) estimates across dose levels. At the MTD dose of 1.0 mg/day, t_1/2 was approximately 2 days, and mean accumulation ratio was 2.4-fold at steady state.

Figure 2. PK and PD features of talazoparib. A–D, Mean concentration–time profiles of talazoparib. Linear mean talazoparib plasma concentration–time profiles over the initial 24 hours after dose and log-linear mean talazoparib plasma concentration–time profiles over the complete sampling interval following single and multiple daily dosing of talazoparib. E–H, Dose proportionality of talazoparib PK and dose–response relationships between talazoparib and PBMC PARP activity. E, Plasma C_max following multiple daily doses ranging from 0.025 to 1.1 mg. F, AUC_0–24 following multiple daily doses ranging from 0.025 to 1.1 mg. Filled circles represent the mean value at each dose level, and error bars represent the standard deviations. Solid line represents the power model fit through the data. G, Dose–response relationship between talazoparib and PBMC PARP activity. H, Exposure–response relationship between talazoparib and PBMC PARP activity. Percentage baseline PBMC PARP activity defined as the mean of the predose PARP activity assessments during the multiple dosing assessment phase (i.e., predose assessments on days 15, 22, and 35 of cycle 1). Abbreviations: AUC_0–24, AUC from 0 to 24 h; IC_50, half maximal inhibitory concentration; ID_50, inhibitory dose 50%; PD, pharmacodynamic.
Plasma concentrations, \( C_{\text{max}} \), and area under the plasma concentration–time curve (AUC) estimates increased approximately with doses ranging from 0.025 to 1.1 mg following multiple daily dosing as shown in Fig. 2E–H. Estimates [95% confidence interval (CI)] of the dose proportionality parameter, \( \beta \), for \( C_{\text{max}} \) and AUC from 0 to 24 hours (AUC\(_{0–24}\)) following multiple daily doses of talazoparib were 1.11 (1.01–1.20) and 0.95 (0.84–1.05), respectively.

Results for urinary elimination of the parent compound suggest linear urinary elimination kinetics after daily talazoparib dosing between the 0.025 and 1.1 mg dose levels. Following single doses in Part 1, mean values for the amount of the analyte excreted in urine from 0 to 24 hours (Ae\(_{0–24}\)) and the fraction of urine excretion from 0 to 24 hours (Fe\(_{0–24}\)) generally increased with dose, and average renal clearance from time 0 to 24 hours postdose (ARC\(_{0–24}\)) values were similar across dose levels. Following multiple daily doses in Part 1, Ae\(_{0–24}\) increased with increasing dose, whereas mean Fe\(_{0–24}\) and ARC\(_{0–24}\) values were generally similar across the 0.025 and 1.1 mg/day dose levels.

### Pharmacodynamics

The mean percentage baseline peripheral blood mononuclear cell (PBMC) PARP activities with multiple-dose talazoparib dosing, mg/day

<table>
<thead>
<tr>
<th>PK parameter</th>
<th>0.025 (n = 3)</th>
<th>0.05 (n = 2)</th>
<th>0.1 (n = 2)</th>
<th>0.2 (n = 3)</th>
<th>0.4 (n = 3)</th>
<th>0.6 (n = 3)</th>
<th>0.9 (n = 5)</th>
<th>1.0 (n = 5)</th>
<th>1.1 (n = 7)</th>
</tr>
</thead>
<tbody>
<tr>
<td>( T_{\text{max}, \text{median (min, max)}, h} )</td>
<td>1.02 (0.58.398)</td>
<td>5.43 (17.10)</td>
<td>0.76 (0.75.082)</td>
<td>1.97 (0.10.302)</td>
<td>0.98 (0.75.200)</td>
<td>0.73 (0.598)</td>
<td>1.04 (0.97.207)</td>
<td>0.76 (0.98.200)</td>
<td>1.02 (0.75.200)</td>
</tr>
<tr>
<td>( C_{\text{max}, \text{mean (SD), pg/mL}} )</td>
<td>300 (78.8)</td>
<td>615 (74.2)</td>
<td>1,880 (332)</td>
<td>5,620 (3530)</td>
<td>6,560 (15,000)</td>
<td>11,300 (3,230)</td>
<td>15,400 (7,990)</td>
<td>21,000 (8,410)</td>
<td></td>
</tr>
<tr>
<td>( AUC_{0–24, \text{mean (SD), pg·h/mL}} )</td>
<td>3,960 (759)</td>
<td>9,770 (2,440)</td>
<td>30,000 (4,490)</td>
<td>83,100 (49,300)</td>
<td>6,300 (22,600)</td>
<td>119,000 (19,900)</td>
<td>157,000 (24,500)</td>
<td>202,000 (54,000)</td>
<td></td>
</tr>
<tr>
<td>( t_{1/2, \text{mean (SD), h}} )</td>
<td>107 (84.2)</td>
<td>132 (12.3)</td>
<td>98.2 (4.83)</td>
<td>50.9 (19.1)</td>
<td>90.7 (32.7)</td>
<td>63.7 (12.7)</td>
<td>71.0 (14.5)</td>
<td>50.0 (16.6)</td>
<td>52.8 (23.2)</td>
</tr>
<tr>
<td>( CL_{\text{f, mean (SD), L/h}} )</td>
<td>6.43 (1.23)</td>
<td>5.28 (1.32)</td>
<td>3.37 (1.052)</td>
<td>3.12 (1.911)</td>
<td>3.64 (2.07)</td>
<td>5.23 (0.941)</td>
<td>5.86 (1.39)</td>
<td>5.24 (0.837)</td>
<td></td>
</tr>
<tr>
<td>( V_{\text{f, mean (SD), L}} )</td>
<td>1,180 (791)</td>
<td>1,020 (345)</td>
<td>477 (47.8)</td>
<td>264 (249)</td>
<td>818 (326)</td>
<td>477 (136)</td>
<td>604 (169)</td>
<td>373 (144)</td>
<td>472 (254)</td>
</tr>
<tr>
<td>( C_{\text{min, mean (SD), pg/mL}} )</td>
<td>169 (58.0)</td>
<td>299 (133)</td>
<td>1,020 (107)</td>
<td>2,880 (957)</td>
<td>2,230 (1,050)</td>
<td>3,470 (802)</td>
<td>3,180 (1,590)</td>
<td>3,720 (803)</td>
<td>2,910 (803)</td>
</tr>
</tbody>
</table>

### Abbreviations

- AUC\(_{0–24}\): AUC from 0 to 24 h
- AUC\(_0–24\): AUC from 0 to last quantifiable concentration
- CL\(_{\text{f, mean (SD), L/h}}\): CL/F at steady state; SD, standard deviation; \( T_{\text{max}} \), time to \( C_{\text{max}} \)

Table 3. PK parameters and PARP inhibition following single and multiple daily dosing
by dose level are provided in Table 3 (Supplementary Fig. S1). Overall, PBMC PARP activity decreased with talazoparib dose across the evaluated dose range.

The dose–response and concentration–response relationships between talazoparib and PBMC PARP activity are shown in Fig. 2E–H, and maximum inhibitory effect model parameter estimates are provided in Supplementary Table S1. In the exposure–response curve, an estimated half maximal inhibitory concentration of AUC\(_{0–24}\) was 19,000 pg·h/mL.

**Efficacy**

In 14 patients with breast cancer (all with deleterious BRCA1/2 mutations) treated with talazoparib at 1.0 mg/day, the objective response rate (ORR) was 50% and included one complete response (CR; Table 4). Five patients had stable disease (SD) lasting at least 24 weeks, resulting in a clinical benefit rate (CBR) of 86% for at least 24 weeks. Median progression-free survival (PFS) was 34.6 weeks (95% CI, 27.1–54.0; Table 4). For the total of 18 patients with breast cancer whose tumors carried the BRCA1 mutation, although the BRCA2 mutations treated at any talazoparib dose level, the ORR and CBR were higher in patients whose tumors carried the BRCA2 mutation (ORR, 55%; 6/11 patients; CBR, 91%, 10/11 patients) compared with those who had the BRCA1 mutation (ORR, 38%; 3/8 patients; CBR, 50%, 4/8 patients; percentage change in target lesion size summarized in Fig. 3A). Of note, 1 patient had aberrations in both BRCA1 and BRCA2, although the BRCA2 aberration detected may not be deleterious (Y3098X). Interestingly, in the patients with BRCA1-mutated breast cancer, higher anti-tumor activity was observed in patients with non–triple-negative breast cancer (n = 9) than in those with triple-negative disease (n = 9; CBR, 89% vs. 56% ≥24 weeks; median PFS, 38.3 weeks (95% CI, 2.6–67.4) vs. 20.4 weeks (95% CI, 3.1–36.1)]. Six of the 18 patients with BRCA1-mutated breast cancer had received prior platinum therapy, of whom 2 had an objective response.

In 12 patients with ovarian cancer with deleterious germline BRCA1/2 mutations with measurable disease treated with talazoparib 1.0 mg/day, ORR and CBR lasting at least 24 weeks equaled 42% and 67%, respectively, with a median PFS of 36.4 weeks (Table 4). For all patients with BRCA1-mutated ovarian cancer treated at any talazoparib dose level with measurable disease (n = 25), ORR and CBR lasting at least 24 weeks were 48% (including one CR) and 76%, respectively (percentage change in target lesion size is summarized in Fig. 3B). All 25 patients had received prior platinum-based chemotherapy; the ORR in platinum-sensitive patients was 55% (11/20 patients) compared with 20% (1/5 patients) in platinum-resistant patients.

All 23 patients with SCLC were enrolled in Part 2 and treated with 1.0 mg/day. Median number of prior regimens was 1, ranging from 0 to 2. Two patients had a partial response (PR; ORR, 9%, with duration of response 12.0 and 15.3 weeks, respectively), and a further 4 had SD lasting at least 16 weeks (CBR, 26% ≥16 weeks; Table 4). For the 2 patients with an objective response, both had had an objective response to the last prior platinum therapy, with a platinum-free interval of 6 months or less. Median PFS for this group was 11.1 weeks (95% CI, 4.3–13.0).

Of the 13 patients with pancreatic cancer from Part 1 and Part 2, 4 had clinical benefit (CBR, 31% ≥16 weeks): Two patients had a PR, 1 with BRCA2 mutation, the other with a PALB2 mutation (Table 4). For patients with Ewing sarcoma, no objective response was observed, and the CBR (SD ≥16 weeks) was 23%.

For the 7 patients currently receiving talazoparib on the study as of the data cutoff of March 31, 2015, 4 have ovarian cancer (continuing on study for 27.4, 28.1, 31.5, and 36.6 months), and 1 patient each has breast, pancreatic, or prostate cancer (24.2, 22.8, and 8.4 months, respectively). The starting dose for these patients ranged between 0.9 and 1.0 mg/day; current dose is between 0.5 and 1.0 mg/day.

**DISCUSSION**

Talazoparib is a potent oral PARP1/2 inhibitor that has equivalent catalytic activity to olaparib and rucaparib, but...
Talazoparib demonstrated favorable PK properties with good oral bioavailability, rapid absorption, and dose-proportional increases in total exposure (AUC) over a wide dose range (0.025–1.1 mg/day). Steady state was reached approximately 2 weeks after initiation of daily dosing. Linear urinary elimination kinetics were reported with daily dosing. At the recommended phase II dose of 1.0 mg/day, the t1/2 was approximately 2 days upon multiple dosing; trough talazoparib plasma concentrations were maintained above 10 nmol/L, suggesting that systemic concentrations of talazoparib are sufficient to inhibit PARP activity.

In pharmacodynamic (PD) testing, talazoparib demonstrated PARP inhibition in PBMCs over a relatively wide range of doses. For doses at and above 0.6 mg/day, PARP activity was consistently inhibited in all patients evaluated. PD results suggest that effective PARP inhibition could still be achieved at reduced dose levels.

Talazoparib demonstrated promising antitumor activity in patients with heavily pretreated breast and ovarian cancers associated with deleterious germline BRCA1/2 mutations. Single-agent activity in patients with advanced breast cancer (including patients with triple-negative disease) equaled 50% (ORR) and 86% (CBR). Similarly, in the 12 patients with BRCA-mutated ovarian cancer treated with 1.0 mg/day of talazoparib, ORR and CBR equaled 42% and 67%, respectively.

Of note, 1 responding patient with pancreatic cancer harbored a PALB2 mutation (21); as this mutation is known to recruit BRCA2 and RAD51 to DNA breaks, such findings support a trial in a broader population (those with additional DNA repair deficiencies as opposed to BRCA mutations only), potentially expanding applications for PARP inhibitor therapy.

In conclusion, the findings from this study demonstrate the effectiveness of single-agent talazoparib for the treatment of patients with and without germline BRCA1/2 mutations in ovarian, breast, small cell lung, and pancreatic cancers. Talazoparib has a tolerable safety profile in multiple patients seen over a treatment period exceeding 2 years. The PK properties of talazoparib support once-daily dosing. Data from this phase I trial support a role for talazoparib in the treatment of patients with advanced tumors (inherited and sporadic cancers with DNA repair deficiencies). Talazoparib is currently undergoing further clinical investigation against multiple tumor types, including a phase III trial in patients with metastatic breast cancer with a deleterious BRCA1/2 mutation.

**METHODS**

**Study Design and Participants**

We undertook a phase I study of talazoparib in patients with advanced solid tumors and either germline BRCA1/2 mutations or a strong preclinical rationale for use of a PARP inhibitor. Eligible patients were age 18 years or older and had histologically or cytologically documented unresectable, locally advanced, or metastatic solid tumors not suitable for established therapy or for which standard therapy had failed; Eastern Cooperative Oncology Group Performance Status of 0 or 1; and adequate hematologic and liver function.
Patients enrolled in Part 1 (dose escalation) had tumors known to harbor DNA repair deficiencies (Supplementary Methods); provision of documentation (genomic or immunohistochemistry) was not required. Enrollment in Part 2 was restricted to patients with selected tumors with confirmed BRCA1/2 germline pathogenic or deleterious mutations by BRACAnalysis (Myriad Genetics) or local laboratory evaluation (ovarian or peritoneal, breast, prostate, or pancreatic cancers), patients with DNA repair deficiency, or patients with SCLC or Ewing sarcoma (Supplementary Methods). Patient eligibility, including a full list of exclusion criteria, is provided in Supplementary Methods.

The study was conducted in accordance with the protocol, good clinical practice standards, and the Declaration of Helsinki and the International Conference on Harmonisation. The appropriate Institutional Review Board or ethics committee at each participating institution approved the protocol. All enrolled patients provided written informed consent before undergoing study-specific procedures.

Study Treatment

For Part 1, fasted patients received a single dose of talazoparib at the start of the study and then underwent PK and PD assessments 1 week later. Following assessments, patients received talazoparib once daily, continuously for 28 days, again followed by a 1-week break from treatment (defined as cycle 1) to assess PK and PD. Dosing was continuous thereafter without breaks except as needed for toxicity. A standard 3+3 design was used for dose escalation (22), with a starting talazoparib dose of 0.025 mg/day. Dose doubling occurred provided toxicities were Common Terminology Criteria for Adverse Events grade 1 or less during cycle 1; dose escalations were limited to 25% to 50% once grade 2 drug-related toxicities were observed (25% for grade 3 drug-related toxicity). For each cohort, the first patient was observed for 15 days prior to additional patient enrollment. To be eligible for DLT assessment, a patient must have received at least 24 of the planned 28 doses of talazoparib between days 8 and 35. A DLT was defined as any of the following events occurring during cycle 1: grade 4 neutropenia associated with grade 2 or greater infection or lasting at least 5 days; grade 4 thrombocytopenia (or grade 3 with either hemorrhage or dose interruption for ≥5 days); any AE of grade 3 or greater considered related to talazoparib, except a nonhematologic asymptomatic grade 3 laboratory AE, grade 3 nausea, vomiting, and/or diarrhea medically managed to grade 2 or less within 24 hours, or grade 3 fatigue that improved to grade 2 or less in no more than 5 days (additional information provided in Supplementary Methods).

Enrollment in Part 2 proceeded once the MTD was determined. Patients received talazoparib at the MTD of 1.0 mg/day starting from cycle 1, day 1 (28-day cycles). Participation in the study could be discontinued at any time at the discretion of the investigator and in accordance with clinical judgment. AEs were recorded from the time of first dose of talazoparib until 30 days after the last dose.

Study Procedures

At screening, patients underwent physical examination (with vital signs and performance status assessment). Safety laboratory tests (complete blood count with differential and platelets, chemistry) were obtained weekly; coagulation and urinalysis were obtained weekly (cycle 1) and at the beginning of each cycle thereafter. Hematology evaluations were conducted more frequently upon observation of grade 2 or greater neutropenia or thrombocytopenia. Further details of study procedures are given in Supplementary Methods.

Pharmacokinetic Analysis

Plasma and urine samples were assayed for talazoparib concentrations using a validated high-performance LC/MS-MS detection method. For plasma, the lower limit of quantitation (LLOQ) was 5.0 pg/mL; for urine, the LLOQ was 25.0 pg/mL. Talazoparib PK parameters (following single and multiple daily dosing) were obtained using standard noncompartmental analysis methods in Phoenix WinNonlin Version 6.4 (Certara L.P.). PK parameters estimated included Cmax; time to Cmax; AUC0–24, AUC from time 0 to time of last quantifiable concentration, and AUC from time 0 extrapolated to infinity; CL/F; V/F; and t1/2. The multiple-dose PK parameters also estimated included minimum plasma concentration and CL/F at steady state. Dose proportionality following single and multiple daily dosing of talazoparib was assessed using a power model approach (23).

Pharmacodynamic Analysis

See Supplementary Methods for details.

Statistical Analysis

The primary objective in Part 1 of this study was to determine the MTD and recommended dose of daily oral talazoparib, secondary objectives included safety, PK, and PD profiles. For Part 2, efficacy parameters in the selected tumor types were investigated per a prespecified analysis based on RECIST version 1.1 through investigator assessment of lesion measurements, including ORR (in patients with measurable disease) or disease-specific changes in tumor markers using standard definitions (24–26). The number and percentage of patients achieving a response were summarized with an exact 95% CI calculated using the Clopper-Pearson method. The PFS was summarized using the Kaplan-Meier method. The data cutoff was March 31, 2015. SAS Analytics Software (version 9.1; SAS Institute, Inc.) was used for data analyses.

Disclosure of Potential Conflicts of Interest

J. de Bono reports receiving honoraria from the Speakers Bureau of AstraZeneca, Genentech, GSK, Merck, and Pfizer, and is a consultant/advisory board member for AstraZeneca, Genentech, GSK, Merck, and Pfizer. R.K. Ramanathan has received travel reimbursement from BioMarin. R. Chugh reports receiving commercial research grants from BioMarin and Medivation. S. Kaye is a consultant/advisory board member for AstraZeneca. J. Heymach is a consultant/advisory board member for BioMarin. N.J. Curtin reports receiving a commercial research grant from BioMarin and commercial research support from Pfizer; has ownership interest (including patents) in Newcastle University and Pfizer; and is a consultant/advisory board member for AbbVie and Tesaro. Z.A. Wainberg is a consultant/advisory board member for Medivation. No potential conflicts of interest were disclosed by the other authors.

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