Supplementary Method (for Supplementary Figures S4D-S4F)

Immunofluorescence analysis of macrophage polarization

Raw264.7 cells, freshly-isolated primary mouse macrophages, or primary mouse macrophages after polarization were fixed in 4% paraformaldehyde-PBS for 10 minutes at RT. Cells then were permeabilized with 0.2 % Triton X-100 in PBS for 10 minutes at RT and incubated with protein block serum-free reagent (Dako) for 1 hour at RT. Primary antibodies (anti-F4/80 at 1:250; anti-iNOS at 1:100; anti-Ym1 at 1:200) were incubated overnight at 4 °C. Cells were then washed with PBS containing 0.05 % Tween-20 (PBST) and incubated with indicated 2^{nd} antibody (AlexaFluor-488 or AlexaFluor-568, at 1:500) for 1 hour at RT. After vigorous washes with PBST, cells were mounted using PermaFluor mounting media (Thermo Scientific). Images were taken using an Olympus IX71 fluorescent microscope with a 10x objective and a DP70 digital camera and processed using Photoshop software.